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•										yrd			CAG Gln		48
													GCT Ala		96
													AGC Ser		144
													TGG Trp		192
													lys	GCC	240
	Cys					Ala								SCT Ala 55	286
					Val					Pro				lys	336
				Pro					Pro				Tyr	Lys	384
			Gly					Lys				Gly		AGC Ser	
		Glu					1	TATO	TGC	CTGC	GGG				470

# (57) Abstract

Purified BMP-9 proteins and processes for producing them are disclosed. The proteins may be used in the treatment of bone and cartilage defects and in wound healing and related tissue repair.

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# BMP-9 COMPOSITIONS

The present invention relates to a novel family of purified proteins designated BMP-9 proteins and processes for obtaining them. These proteins may be used to induce bone and/or cartilage formation and in wound healing and tissue repair.

The murine BMP-9 DNA sequence (SEQ ID NO: 1) and amino acid sequence (SEQ ID NO: 2) are set forth in Figure 1. Human BMP-9 sequence is set forth in Figure 3 (SEQ ID NO: 8 and SEQ ID NO: 9). It is contemplated that BMP-9 proteins are capable of inducing the formation of cartilage and/or bone. BMP-9 proteins may be further characterized by the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay described below.

Murine BMP-9 is characterized by comprising amino acid #319 to #428 of Figure 1 (SEQ ID NO: 2 amino acids #1-110). Murine BMP-9 may be produced by culturing a cell transformed with a DNA sequence comprising nucleotide #610 to nucleotide #1893 as shown in Figure 1 (SEQ ID NO: 1) and recovering and purifying from the culture medium a protein characterized by the amino acid sequence comprising amino acid #319 to #428 as shown in Figure 1 (SEQ ID NO: 2) substantially free from other proteinaceous materials with which it is co-produced.

Human BMP-9 is expected to be homologous to murine BMP-9 and is characterized by comprising amino acid #1 (Ser, Ala, Gly) to #110 of Figure 3 (SEQ ID NO: 9) (Arg). The invention includes methods for obtaining the DNA sequences encoding human BMP-9. This method entails utilizing the murine BMP-9 nucleotide sequence or portions thereof to design probes to screen libraries for the human gene or fragments thereof using standard techniques. Human BMP-9 may be produced by culturing

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a cell transformed with the BMP-9 DNA sequence and recovering and purifying BMP-9 from the culture medium. The expressed protein is isolated, recovered, and purified from the culture medium. The purified expressed protein is substantially free from other proteinaceous materials with which it is coproduced, as well as from other contaminants. The recovered purified protein is contemplated to exhibit cartilage and/or bone formation activity. The proteins of the invention may be further characterized by the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay described below.

Human BMP-9 may be produced by culturing a cell transformed with a DNA sequence comprising nucleotide #124 to #453 as shown in SEQ ID NO: 8 and recovering and purifying from the culture medium a protein characterized by the amino acid sequence of SEQ ID NO: 9 from amino acid #1 to amino acid #110 substantially free from other proteinaceous materials with which it is co-produced.

Another aspect of the invention provides pharmaceutical compositions containing a therapeutically effective amount of a BMP-9 protein in a pharmaceutically acceptable vehicle or carrier. BMP-9 compositions of the invention may be used in the formation of cartilage. These compositions may further be utilized for the formation of bone. BMP-9 compositions may also be used for wound healing and tissue repair. Compositions of the invention may further include at least one other therapeutically useful agent such as the BMP proteins BMP-1, BMP-2, BMP-3, BMP-4, BMP-5, BMP-6, and BMP-7 disclosed for instance in PCT publications WO88/00205, WO89/10409, and WO90/11366, and BMP-8, disclosed in U.S. application Ser. No. 07/641,204 filed January 15, 1991, Ser. No. 07/525,357 filed May 16, 1990, and Ser. No. 07/800,364 filed November 20, 1991.

The compositions of the invention may comprise, in addition to a BMP-9 protein, other therapeutically useful agents including growth factors such as epidermal growth factor (EGF), fibroblast growth factor (FGF), transforming growth

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factor (TGF- $\alpha$  and TGF- $\beta$ ), and insulin-like growth factor (IGF). The compositions may also include an appropriate matrix for instance, for supporting the composition and providing a surface for bone and/or cartilage growth. The matrix may provide slow release of the osteoinductive protein and/or the appropriate environment for presentation thereof.

The BMP-9 compositions may be employed in methods for treating a number of bone and/or cartilage defects, periodontal disease and various types of wounds. These methods, according to the invention, entail administering to a patient needing such bone and/or cartilage formation wound healing or tissue repair, an effective amount of a BMP-9 protein. These methods may also entail the administration of a protein of the invention in conjunction with at least one of the novel BMP proteins disclosed in the co-owned applications described above. In addition, these methods may also include the administration of a BMP-9 protein with other growth factors including EGF, FGF, TGF- $\alpha$ , TGF- $\beta$ , and IGF.

Still a further aspect of the invention are DNA sequences coding for expression of a BMP-9, protein. Such sequences include the sequence of nucleotides in a 5' to 3' direction illustrated in Figure 1 (SEQ ID NO: 1) and Figure 3 (SEQ ID NO: 8) or DNA sequences which hybridize under stringent conditions with the DNA sequences of Figure 1 or 3 and encode a protein having the ability to induce the formation of cartilage and/or bone. Finally, allelic or other variations of the sequences of Figure 1 or 3, whether such nucleotide changes result in changes in the peptide sequence or not, are also included in the present invention.

A further aspect of the invention includes vectors comprising a DNA sequence as described above in operative association with an expression control sequence therefor. These vectors may be employed in a novel process for producing a BMP-9 protein of the invention in which a cell line transformed with a DNA sequence encoding a BMP-9 protein in

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operative association with an expression control sequence therefor, is cultured in a suitable culture medium and a BMP-9 protein is recovered and purified therefrom. This process may employ a number of known cells both prokaryotic and eukaryotic as host cells for expression of the polypeptide.

Other aspects and advantages of the present invention will be apparent upon consideration of the following detailed description and preferred embodiments thereof.

# 10 Brief Description of the Drawing

FIG. 1 comprises DNA sequence and derived amino acid sequence of murine BMP-9 from clone ML14a further described below.

FIG. 2 comprises DNA sequence and derived amino acid sequence of human BMP-4 from lambda U2OS-3 ATCC #40342.

FIG. 3 comprises DNA sequence and derived amino acid sequence of human BMP-9 from  $\lambda$  FIX/H6111 ATCC # 75252.

# 20 <u>Detailed Descripton of the Invention</u>

The murine BMP-9 nucleotide sequence (SEQ ID NO: 1) and encoded amino acid sequence (SEQ ID NO: 2) are depicted in Figure 1. Purified murine BMP-9 proteins of the present invention are produced by culturing a host cell transformed wth a DNA sequence comprising the DNA coding sequence of Figure 1 (SEQ ID NO: 1) from nucleotide #610 to nucleotide #1893 and recovering and purifying from the culture medium a protein which contains the amino acid sequence or a substantially homologous sequence as represented by amino acid #319 to #428 of Figure 1 (SEQ ID NO: 2). The BMP-9 proteins recovered from the culture medium are purified by isolating them from other proteinaceous materials from which they are co-produced and from other contaminants present.

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Human BMP-9 nucleotide and amino acid sequence is depicted in SEQ ID No: 8 and 9. Mature human BMP-9 is expected to comprise amino acid #1 (Ser, Ala, Gly) to #110 (Arg).

Human BMP-9 may be produced by culturing a cell transformed with a DNA sequence comprising nucleotide #124 to #453 as shown in SEQ ID NO: 8 and recovering and purifying from the culture medium a protein characterized by the amino acid sequence of SEQ ID NO: 9 from amino acid #1 to amino acid #110 substantially free from other proteinaceous materials with which it is co-produced.

BMP-9 proteins may be characterized by the ability to induce the formation of cartilage. BMP-9 proteins may be further characterized by the ability to induce the formation of bone. BMP-9 proteins may be further characterized by the ability to demonstrate cartilage and/or bone formation activity in the rat bone formation assay described below.

The BMP-9 proteins provided herein also include factors encoded by the sequences similar to those of Figure 1 and 3 (SEQ ID NO's: 1 and 8), but into which modifications are naturally provided (e.g. allelic variations in the nucleotide sequence which may result in amino acid changes in the polypeptide) or deliberately engineered. For example, synthetic polypeptides may wholly or partially duplicate continuous sequences of the amino acid residues of Figure 1 of Figure 3 (SEQ ID NO's: 2 and 9). These sequences, by virtue of sharing primary, secondary, or tertiary structural conformational characteristics with bone growth factor polypeptides of Figure 1 and Figure 3 may possess bone growth factor biological properties in common therewith. may be employed as biologically active substitutes for naturally-occurring BMP-9 and other BMP-9 polypeptides in therapeutic processes.

Other specific mutations of the sequences of BMP-9 proteins described herein involve modifications of glycosylation sites. These modifications may involve O-linked

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or N-linked glycosylation sites. For instance, the absence of glycosylation or only partial glycosylation results from amino deletion at asparagine-linked substitution or glycosylation recognition sites. The asparagine-linked glycosylation recognition sites comprise tripeptide sequences which are specifically recognized by appropriate cellular glycosylation enzymes. These tripeptide sequences are either asparagine-X-threonine or asparagine-X-serine, where X is usually any amino acid. A variety of amino acid substitutions or deletions at one or both of the first or third amino acid positions of a glycosylation recognition site (and/or amino acid deletion at the second position) results in nonglycosylation at the modified tripeptide sequence.

The present invention also encompasses the novel DNA sequences, free of association with DNA sequences encoding other proteinaceous materials, and coding on expression for BMP-9 proteins. These DNA sequences include those depicted in Figure 1 or Figure 3 (SEQ ID NO's: 1 and 8) in a 5' to 3' direction and those sequences which hybridize thereto under stringent hybridization conditions [see, T. Maniatis et al, Molecular Cloning (A Laboratory Manual), Cold Spring Harbor Laboratory (1982), pages 387 to 389] and encode a protein having cartilage and/or bone inducing activity.

Similarly, DNA sequences which code for BMP-9 proteins coded for by the sequences of Figure 1 or Figure 3, but which differ in codon sequence due to the degeneracies of the genetic code or allelic variations (naturally-occurring base changes in the species population which may or may not result in an amino acid change) also encode the novel factors described herein. Variations in the DNA sequences of Figure 1 or Figure 3 (SEQ ID NO: 1 and 8) which are caused by point mutations or by induced modifications (including insertion, deletion, and substitution) to enhance the activity, half-life or production of the polypeptides encoded are also encompassed in the invention.

Another aspect of the present invention provides a novel

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method for producing BMP-9 proteins. The method of the present invention involves culturing a suitable cell line, which has been transformed with a DNA sequence encoding a BMP-9 protein of the invention, under the control of known regulatory sequences. The transformed host cells are cultured and the BMP-9 proteins recovered and purified from the culture medium. The purified proteins are substantially free from other proteins with which they are co-produced as well as from other contaminants.

Suitable cells or cell lines may be mammalian cells, such as Chinese hamster ovary cells (CHO). The selection of suitable mammalian host cells and methods for transformation, culture, amplification, screening, product production and purification are known in the art. See, e.g., Gething and Sambrook, Nature, 293:620-625 (1981), or alternatively, Kaufman et al, Mol. Cell. Biol., 5(7):1750-1759 (1985) or Howley et al, U.S. Patent 4,419,446. Another suitable mammalian cell line, which is described in the accompanying examples, is the monkey COS-1 cell line. The mammalian cell CV-1 may also be suitable.

Bacterial cells may also be suitable hosts. For example, the various strains of  $\underline{E}$ .  $\underline{coli}$  (e.g., HB101, MC1061) are well-known as host cells in the field of biotechnology. Various strains of  $\underline{B}$ .  $\underline{subtilis}$ ,  $\underline{Pseudomonas}$ , other bacilli and the like may also be employed in this method.

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Many strains of yeast cells known to those skilled in the art may also be available as host cells for expression of the polypeptides of the present invention. Additionally, where desired, insect cells may be utilized as host cells in the method of the present invention. See, e.g. Miller et al, Genetic Engineering, 8:277-298 (Plenum Press 1986) and references cited therein.

Another aspect of the present invention provides vectors for use in the method of expression of these novel BMP-9 polypeptides. Preferably the vectors contain the full novel

DNA sequences described above which encode the novel factors of Additionally the vectors also contain invention. appropriate expression control sequences permitting expression of the BMP-9 protein sequences. Alternatively, vectors incorporating modified sequences as described above are also embodiments of the present invention. The vectors may be employed in the method of transforming cell lines and contain selected regulatory sequences in operative association with the DNA coding sequences of the invention which are capable of directing the replication and expression thereof in selected host cells. Regulatory sequences for such vectors are known to those skilled in the art and may be selected depending upon the host cells. Such selection is routine and does not form part of the present invention.

protein of the present invention, which induces cartilage and/or bone formation in circumstances where bone is not normally formed, has application in the healing of bone fractures and cartilage defects in humans and other animals. Such a preparation employing a BMP-9 protein may have prophylactic use in closed as well as open fracture reduction and also in the improved fixation of artificial joints. novo bone formation induced by an osteogenic agent contributes to the repair of congenital, trauma induced, or oncologic resection induced craniofacial defects, and also is useful in cosmetic plastic surgery. A BMP-9 protein may be used in the treatment of periodontal disease, and in other tooth repair processes. Such agents may provide an environment to attract bone-forming cells, stimulate growth of bone-forming cells or induce differentiation of progenitors of bone-forming cells. BMP-9 polypeptides of the invention may also be useful in the treatment of osteoporosis. A variety of osteogenic, cartilage-inducing and bone inducing factors have been described. See, e.g. European patent applications 148;155 and 169,016 for discussions thereof.

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The proteins of the invention may also be used in wound healing and related tissue repair. The types of wounds include, but are not limited to burns, incisions and ulcers. (See, e.g. PCT Publication WO84/01106 for discussion of wound healing and related tissue repair).

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It is further contemplated that proteins of the invention may increase neuronal survival and therefore be useful in transplantation and treatment of conditions exhibiting a decrease in neuronal survival.

A further aspect of the invention is a therapeutic method and composition for repairing fractures and other conditions related to cartilage and/or bone defects or periodontal diseases. The invention further comprises therapeutic methods and compositions for wound healing and tissue repair. Such compositions comprise a therapeutically effective amount of at least one of the BMP-9 proteins of the invention in admixture with a pharmaceutically acceptable vehicle, carrier or matrix.

It is expected that the proteins of the invention may act in concert with or perhaps synergistically with other related proteins and growth factors. Further therapeutic methods and compositions of the invention therefore comprise a therapeutic amount of at least one BMP-9 protein of the invention with a therapeutic amount of at least one of the other BMP proteins disclosed in co-owned applications described above. combinations may comprise separate molecules of the BMP proteins or heteromolecules comprised of different For example, a method and composition of the moieties. invention may comprise a disulfide linked dimer comprising a BMP-9 protein subunit and a subunit from one of the "BMP" proteins described above. A further embodiment may comprise a heterodimer of BMP-9 moieties. Further, BMP-9 proteins may be combined with other agents beneficial to the treatment of the bone and/or cartilage defect, wound, or tissue in question. These agents include various growth factors such as epidermal growth factor (EGF), platelet derived growth factor (PDGF),

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transforming growth factors (TGF- $\alpha$  and TGF- $\beta$ ), and insulin-like growth factor (IGF).

The preparation and formulation of such physiologically acceptable protein compositions, having due regard to pH, isotonicity, stability and the like, is within the skill of the art. The therapeutic compositions are also presently valuable for veterinary applications due to the lack of species specificity in BMP proteins. Particularly domestic animals and thoroughbred horses in addition to humans are desired patients for such treatment with BMP-9 of the present invention.

therapeutic method includes administering composition topically, systemically, or locally as an implant or device. When administered, the therapeutic composition for use in this invention is, of course, in a pyrogen-free, physiologically acceptable form. Further, the composition may desirably be encapsulated or injected in a viscous form for delivery to the site of bone, cartilage or tissue damage. Topical administration may be suitable for wound healing and tissue repair. Therapeutically useful agents other than the BMP-9 proteins which may also optionally be included in the above, composition as described may alternatively additionally, be administered simultaneously or sequentially with the BMP composition in the methods of the invention.

Preferably for bone and/or cartilage formation, the composition would include a matrix capable of delivering BMP-9 or other BMP proteins to the site of bone and/or cartilage damage, providing a structure for the developing bone and cartilage and optimally capable of being resorbed into the body. The matrix may provide slow release of BMP-9 and/or the appropriate environment for presentation thereof. Such matrices may be formed of materials presently in use for other implanted medical applications.

The choice of matrix material is based on biocompatibility, biodegradability, mechanical properties, cosmetic appearance and interface properties. The particular

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application of the BMP-9 compositions will define appropriate formulation. Potential matrices for the compositions may be biodegradable and chemically defined sulfate, tricalciumphosphate, hydroxyapatite, polylactic acid and polyanhydrides. Other potential materials are biodegradable and biologically well defined, such as bone Further matrices are comprised of pure or dermal collagen. proteins or extracellular matrix components. Other potential matrices are nonbiodegradable and chemically defined, such as sintered hydroxyapatite, bioglass, aluminates, or other ceramics. Matrices may be comprised of combinations of any of the above mentioned types of material, such as polylactic acid and hydroxyapatite or collagen and tricalciumphosphate. bioceramics may be altered in composition, such as in calciumaluminate-phosphate and processing to alter pore size, particle size, particle shape, and biodegradability.

The dosage regimen will be determined by the attending physician considering various factors which modify the action of the BMP-9 protein, e.g. amount of bone weight desired to be formed, the site of bone damage, the condition of the damaged bone, the size of a wound, type of damaged tissue, the patient's age, sex, and diet, the severity of any infection, time of administration and other clinical factors. The dosage may vary with the type of matrix used in the reconstitution and the types of BMP proteins in the composition. The addition of other known growth factors, such as IGF I (insulin like growth factor I), to the final composition, may also effect the Progress can be monitored by periodic assessment of dosage. for example, x-rays, and/or repair, bone growth histomorphometric determinations and tetracycline labeling.

The following examples illustrate practice of the present invention in recovering and characterizing murine BMP-9 protein and employing it to recover the human and other BMP-9 proteins, obtaining the human proteins and expressing the proteins via recombinant techniques.

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# EXAMPLE I Murine BMP-9

750,000 recombinants of a mouse liver cDNA library made in the vector lambdaZAP (Stratagene/Catalog #935302) are plated and duplicate nitrocellulose replicas made. A fragment of human BMP-4 DNA corresponding to nucleotides 1330-1627 of Figure 2 (SEQ ID NO: 3) (the human BMP-4 sequence) is "Plabeled by the random priming procedure of Feinberg et al. [Anal. Biochem. 132: 6-13 (1983)] and hybridized to both sets of filters in SHB at 60°C for 2 to 3 days. Both sets of filters are washed under reduced stringency conditions (4X SSC, 0.1% Many duplicate hybridizing recombinants of SDS at 60°C). various intensities (approximately 92) are noted. 50 of the strongest hybridizing recombinant bacteriophage are plaque purified and their inserts are transferred to the plasmid Bluescript SK (+/-) according to the in vivo excision protocol described by the manufacturer (Stratagene). analysis of several recombinants indicate that they encode a protein homologous to other BMP proteins and other proteins in the TGF- $\beta$  family. The DNA sequence and derived amino acid sequence of one recombinant, designated ML14a, is set forth in Figure 1. (SEQ ID NO: 1)

The nucleotide sequence of clone ML14a contains an open reading frame of 1284 bp, encoding a BMP-9 protein of 428 amino acids. The encoded 428 amino acid BMP-9 protein is contemplated to be the primary translation product as the coding sequence is preceded by 609 bp of 5' untranslated sequence with stop codons in all three reading frames. The 428 amino acid sequence predicts a BMP-9 protein with a molecular weight of 48,000 daltons.

Based on knowledge of other BMP proteins and other proteins within the TGF- $\beta$  family, it is predicted that the precursor polypeptide would be cleaved at the multibasic sequence ARG-ARG-LYS-ARG in agreement with a proposed consensus

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proteolytic processing sequence of ARG-X-X-ARG. Cleavage of the BMP-9 precursor polypeptide at this location would generate a 110 amino acid mature peptide beginning with the amino acid SER at position #319. The processing of BMP-9 into the mature form is expected to involve dimerization and removal of the N-terminal region in a manner analogous to the processing of the related protein TGF- $\beta$  [L.E. Gentry, et al., Molec. & Cell. Biol. 8:4162 (1988); R. Derynck, et al., Nature 316:701 (1985)].

It is contemplated therefore that the mature active species of murine BMP-9 comprises a homodimer of 2 polypeptide subunits, each subunit comprising amino acids #319-#428 with a predicted molecular weight of approximately 12,000 daltons. Further active species are contemplated comprising amino acids #326 - #428 thereby including the first conserved cysteine residue. As with other members of the BMP and TGF- $\beta$  family of proteins, the carboxy-terminal region of the BMP-9 protein exhibits greater sequence conservation than the more aminoterminal portion. The percent amino acid identity of the murine BMP-9 protein in the cysteine-rich C-terminal domain (amino acids #326 - #428) to the corresponding region of other human BMP proteins and other proteins within the TGF- $\beta$  family is as follows: BMP-2, 53%; BMP-3, 43%; BMP-4, 53%; BMP-5, 55%; BMP-6, 55%; BMP-7, 53%; Vgl, 50%; GDF-1, 43%; TGF- $\beta$ 1, 32%; TGF- $\beta$ 2, 34%; TGF- $\beta$ 3, 34%; inhibin  $\beta$ (B), 34%; and inhibin  $\beta$ (A), 42%.

#### EXAMPLE II

#### Human BMP-9

Murine and human osteoinductive factor genes are presumed to be significantly homologous, therefore the murine coding sequence or a portion thereof is used as a probe to screen a human genomic library or as a probe to identify a human cell line or tissue which synthesizes the analogous human cartilage and/or bone protein. A human genomic library (Toole et al., supra) may be screened with such a probe, and presumptive

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positives isolated and DNA sequence obtained. Evidence that this recombinant encodes a portion of the human BMP-9 relies of the murine/human protein and gene structure homologies.

Once a recombinant bacteriophage containing DNA encoding portion of the human cartilage and/or bone inductive factor molecule is obtained, the human coding sequence can be used as a probe to identify a human cell line or tissue which synthesizes BMP-9. Alternatively, the murine coding sequence can be used as a probe to identify such human cell line or tissue. Briefly described, RNA is extracted from a selected cell or tissue source and either electrophoresed on a formaldehyde agarose gel and transferred to nitrocellulose, or reacted with formaldehyde and spotted on nitrocellulose directly. The nitrocellulose is then hybridized to a probe derived from a coding sequence of the murine or human BMP-9. mRNA is selected by oligo (dT) cellulose chromatography and cDNA is synthesized and cloned in lambda gt10 or lambda ZAP by established techniques (Toole et al., supra).

Additional methods known to those skilled in the art may be used to isolate the human and other species' BMP-9 proteins of the invention.

### A. Isolation of Human BMP-9 DNA

One million recombinants of a human genomic library constructed in the vector  $\lambda$ FIX (Stratagene catalog # 944201) are plated and duplicate nitrocellulose replicas made. Two oligonucleotides probes designed on the basis of nucleotides #1665-#1704 and #1837-#1876 of the sequence set forth in Figure 1 (SEQ ID NO:1) are synthesized on an automated DNA synthesizer. The sequence of these two oligonucleotides is indicated below:

#1: CTATGAGTGTAAAGGGGGTTGCTTCTTCCCATTGGCTGAT

#2: GTGCCAACCCTCAAGTACCACTATGAGGGGATGAGTGTGG

These two oligonucleotide probes are radioactively labeled with

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 $\gamma^{12}$ P-ATP and each is hybridized to one set of the duplicate nitrocellulose replicas in SHB at 65°C and washed with 1X SSC, 0.1% SDS at 65°C. Three recombinants which hybridize to both oligonucleotide probes are noted. All three positively hybridizing recombinants are plaque purified, bacteriophage plate stocks are prepared and bacteriophage DNA is isolated from each. The oligonucleotide hybridizing regions of one of these recombinants, designated HG111, is localized to a 1.2 kb Pst I/Xba I fragment. This fragment is subcloned into a plasmid vector (pGEM-3) and DNA sequence analysis is performed. HG111 was deposited with the ATCC, 12301 Parklawn Drive, Rockville, Maryland USA on June 16, 1992 under the requirements of the Budapest Treaty and designated as ATCC # 75252. subclone is designated pGEM-111. A portion of the DNA sequence of clone pGEM-111 is set forth in Figure 3 (SEQ ID NO:8/ HUMAN BMP-9 sequence). This sequence encodes the entire mature region of human BMP-9 and a portion of the propeptide. should be noted that this sequence consists of preliminary Particularly, the propeptide region is subject to data. further analysis and characterization. For example, nucleotides #1 through #3 (TGA) encode a translational stop which may be incorrect due to the preliminary nature of the sequence. It is predicted that additional sequences present in both pGEM-111 (the 1.2 kb PstI/XbaI fragment of HG111 subcloned into pGEM) and HG111 encode additional amino acids of the human BMP-9 propeptide region. Based on knowledge of other BMPs and other proteins within the TGF- $\beta$  family, it is predicted that the precursor polypeptide would be cleaved at the multibasic sequence ARG-ARG-LYS-ARG (amino acids # -4 through # -1 of SEQUENCE ID NO:9) in agreement with a proposed consensus proteolytic processing sequence ARG-X-X-ARG. Cleavage of the human BMP-9 precursor polypeptide at this location would generate a 110 amino acid mature peptide beginning with the amino acid SER at position #1 of SEQUENCE ID NO:9 (encoded by

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nucleotides #124 through #126 of SEQUENCE ID NO:8). The processing of human BMP-9 into the mature form is expected to involve dimerization and removal of the N-terminal region in a manner analogous to the processing of the related protein TGF- $\beta$  [L.E. Gentry, et al., Molec. & Cell. Biol. 8:4162 (1988); R. Derynck, et al., Nature 316:701 (1985)].

It is contemplated therefore that the mature active species of human BMP-9 comprises a homodimer of two polypeptide subunits, each subunit comprising amino acids #1 through #110 of SEQUENCE ID NO:9, with a predicted molecular weight of Further active species are contemplated 12,000 daltons. comprising amino acids #8 through #110 thereby including the first conserved cysteine residue. As with other members of the BMP and TGF- $\beta$  family of proteins, the carboxy-terminal portion of the human BMP-9 sequence exhibits greater sequence conservation than the amino-terminal portion. the percent amino acid identity of the human BMP-9 protein in the cysteinerich C-terminal domain (amino acids #8 through #110) to the corresponding region of other human BMP proteins and other proteins within the TGF- $\beta$  family is as follows: BMP-2, 52%; BMP-3, 40%; BMP-4, 52%; BMP-5, 55%; BMP-6, 55%; BMP-7, 53%; murine BMP-9, 97%; Vq1, 50%; GDF-1, 44%; TGF- $\beta$ 1, 32%; TGF- $\beta$ 2, 32%; TGF- $\beta$ 3, 32%; inhibin  $\beta$  (B), 35%; and inhibin  $\beta$  (A), 41%.

#### 25 EXAMPLE III

# Rosen Modified Sampath-Reddi Assay

A modified version of the rat bone formation assay described in Sampath and Reddi, Proc. Natl. Acad. Sci. U.S.A., 80:6591-6595 (1983) is used to evaluate bone and/or cartilage activity of the BMP proteins. This modified assay is herein called the Rosen-modified Sampath-Reddi assay. The ethanol precipitation step of the Sampath-Reddi procedure is replaced by dialyzing (if the composition is a solution) or diafiltering (if the composition is a suspension) the fraction to be assayed against water. The solution or suspension is then redissolved

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in 0.1 % TFA, and the resulting solution added to 20mg of rat matrix. A mock rat matrix sample not treated with the protein serves as a control. This material is frozen and lyophilized and the resulting powder enclosed in #5 gelatin capsules. The capsules are implanted subcutaneously in the abdominal thoracic area of 21 - 49 day old male Long Evans rats. The implants are removed after 7 - 14 days. Half of each implant is used for alkaline phosphatase analysis [See, A. H. Reddi et al., Proc. Natl Acad Sci., 69:1601 (1972)].

The other half of each implant is fixed and processed for histological analysis. 1μm glycolmethacrylate sections are stained with Von Kossa and acid fuschin to score the amount of induced bone and cartilage formation present in each implant. The terms +1 through +5 represent the area of each histological section of an implant occupied by new bone and/or cartilage cells and matrix. A score of +5 indicates that greater than 50% of the implant is new bone and/or cartilage produced as a direct result of protein in the implant. A score of +4, +3, +2 and +1 would indicate that greater than 40%. 30%, 20% and 10% respectively of the implant contains new cartilage and/or bone. In a modified scoring method, three non-adjacent sections are evaluated from each implant and "+/-" indicates tentative identification of cartilage or bone; "+1" indicates >10% of each section being new cartilage or bone; "+2", >25%; "+3", >50%; "+4", ~75%; "+5", >80%. A "-" indicates that the implant is not recovered.

It is contemplated that the dose response nature of the BMP-9 containing samples of the matrix samples will demonstrate that the amount of bone and/or cartilage formed increases with the amount of BMP-9 in the sample. It is contemplated that the control samples will not result in any bone and/or cartilage formation.

As with other cartilage and/or bone inductive proteins such as the above-mentioned "BMP" proteins, the bone and/or cartilage formed is expected to be physically confined to the

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space occupied by the matrix. Samples are also analyzed by SDS gel electrophoresis and isoelectric focusing followed by autoradiography. The activity is correlated with the protein bands and pI. To estimate the purity of the protein in a particular fraction an extinction coefficient of 1 OD/mg-cm is used as an estimate for protein and the protein is run on SDS PAGE followed by silver staining or radioiodination and autoradiography.

#### 10 EXAMPLE IV

### Expression of BMP-9

In order to produce murine, human or other mammalian BMP-9 proteins, the DNA encoding it is transferred into an appropriate expression vector and introduced into mammalian cells or other preferred eukaryotic or prokaryotic hosts by conventional genetic engineering techniques. The preferred expression system for biologically active recombinant human BMP-9 is contemplated to be stably transformed mammalian cells.

One skilled in the art can construct mammalian expression vectors by employing the sequence of Figure 1 (SEQ ID NO: 1) or Figure 3 (SEQ ID NO: 8), or other DNA sequences encoding BMP-9 proteins or other modified sequences and known vectors, such as pCD [Okayama et al., Mol. Cell Biol., 2:161-170 (1982)], pJL3, pJL4 [Gough et al., EMBO J., 4:645-653 (1985)] and pMT2 CXM.

The mammalian expression vector pMT2 CXM is a derivative of p91023 (b) (Wong et al., Science 228:810-815, 1985) differing from the latter in that it contains the ampicillin resistance gene in place of the tetracycline resistance gene and further contains a XhoI site for insertion of cDNA clones. The functional elements of pMT2 CXM have been described (Kaufman, R.J., 1985, Proc. Natl. Acad. Sci. USA 82:689-693) and include the adenovirus VA genes, the SV40 origin of replication including the 72 bp enhancer, the adenovirus major late promoter including a 5' splice site and the majority of

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the adenovirus tripartite leader sequence present on adenovirus late mRNAs, a 3' splice acceptor site, a DHFR insert, the SV40 early polyadenylation site (SV40), and pBR322 sequences needed for propagation in  $\underline{E}$ .  $\underline{coli}$ .

Plasmid pMT2 CXM is obtained by EcoRI digestion of pMT2-VWF, which has been deposited with the American Type Culture Collection (ATCC), Rockville, MD (USA) under accession number ATCC 67122. EcoRI digestion excises the cDNA insert present in pMT2-VWF, yielding pMT2 in linear form which can be ligated and used to transform <u>E</u>. <u>coli</u> HB 101 or DH-5 to ampicillin resistance. Plasmid pMT2 DNA can be prepared by conventional methods. pMT2 CXM is then constructed using loopout/in mutagenesis [Morinaga, et al., <u>Biotechnology</u> 84: 636 (1984). This removes bases 1075 to 1145 relative to the Hind III site near the SV40 origin of replication and enhancer sequences of pMT2. In addition it inserts the following sequence:

5' PO-CATGGGCAGCTCGAG-3' (SEQ ID NO: 5)

at nucleotide 1145. This sequence contains the recognition site for the restriction endonuclease Xho I. A derivative of pMT2CXM, termed pMT23, contains recognition sites for the restriction endonucleases PstI, Eco RI, SalI and XhoI. Plasmid pMT2 CXM and pMT23 DNA may be prepared by conventional methods.

pEMC2bl derived from pMT21 may also be suitable in practice of the invention. pMT21 is derived from pMT2 which is derived from pMT2-VWF. As described above EcoRI digestion excises the cDNA insert present in pMT-VWF, yielding pMT2 in linear form which can be ligated and used to transform <u>E. Coli</u> HB 101 or DH-5 to ampicillin resistance. Plasmid pMT2 DNA can be prepared by conventional methods.

pMT21 is derived from pMT2 through the following two modifications. First, 76 bp of the 5' untranslated region of the DHFR cDNA including a stretch of 19 G residues from G/C tailing for cDNA cloning is deleted. In this process, a XhoI site is inserted to obtain the following sequence immediately

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upstream from DHFR: 5' - CTGCAGGCGAGCCTGAATTCCTCGAGCCATCATG-3'
PstI Eco RI XhoI

(SEQ ID NO: 6)

Second, a unique ClaI site is introduced by digestion with ECORV and XbaI, treatment with Klenow fragment of DNA polymerase I, and ligation to a ClaI linker (CATCGATG). This deletes a 250 bp segment from the adenovirus associated RNA (VAI) region but does not interfere with VAI RNA gene expression or function. pMT21 is digested with EcoRI and XhoI, and used to derive the vector pEMC2B1.

A portion of the EMCV leader is obtained from pMT2-ECAT1 [S.K. Jung, et al, <u>J. Virol 63</u>:1651-1660 (1989)] by digestion with Eco RI and PstI, resulting in a 2752 bp fragment. This fragment is digested with TaqI yielding an Eco RI-TaqI fragment of 508 bp which is purified by electrophoresis on low melting agarose gel. A 68 bp adapter and its complementary strand are synthesized with a 5' TaqI protruding end and a 3' XhoI protruding end which has the following sequence:

20 5'-<u>CGA</u>GGTTAAAAAACGTCTAGGCCCCCCGAACCACGGGGACGTGGTTTTCCTTT
TaqI

GAAAAACACGATTGC-3'
XhoI (SEQ ID NO: 7)

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This sequence matches the EMC virus leader sequence from nucleotide 763 to 827. It also changes the ATG at position 10 within the EMC virus leader to an ATT and is followed by a XhoI site. A three way ligation of the pMT21 Eco RI-XhoI fragment, the EMC virus EcoRI-TaqI fragment, and the 68 bp

oligonucleotide adapter TaqI-XhoI adapter resulting in the vector  $pEMC2\beta1$ .

This vector contains the SV40 origin of replication and enhancer, the adenovirus major late promoter, a cDNA copy of the majority of the adenovirus tripartite leader sequence, a small hybrid intervening sequence, an SV40 polyadenylation

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signal and the adenovirus VA I gene, DHFR and  $\beta$ -lactamase markers and an EMC sequence, in appropriate relationships to direct the high level expression of the desired cDNA in mammalian cells.

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The construction of vectors may involve modification of the BMP-9 DNA sequences. For instance, BMP-9 cDNA can be modified by removing the non-coding nucleotides on the 5' and 3' ends of the coding region. The deleted non-coding nucleotides may or may not be replaced by other sequences known to be beneficial for expression. These vectors are transformed into appropriate host cells for expression of BMP-9 proteins.

One skilled in the art can manipulate the sequences of Figure 1 or Figure 3 (SEQ ID NO: 1 and 8) by eliminating or replacing the mammalian regulatory sequences flanking the coding sequence with bacterial sequences to create bacterial vectors for intracellular or extracellular expression by bacterial cells. For example, the coding sequences could be further manipulated (e.g. ligated to other known linkers or modified by deleting non-coding sequences therefrom or altering nucleotides therein by other known.techniques). The modified BMP-9 coding sequence could then be inserted into a known bacterial vector using procedures such as described in T. Taniguchi et al., Proc. Natl Acad. Sci. USA, 77:5230-5233 This exemplary bacterial vector could then be transformed into bacterial host cells and a BMP-9 protein expressed thereby. For a strategy for producing extracellular expression of BMP-9 proteins in bacterial cells, see, e.g. European patent application EPA 177,343.

Similar manipulations can be performed for the construction of an insect vector [See, e.g. procedures described in published European patent application 155,476] for expression in insect cells. A yeast vector could also be constructed employing yeast regulatory sequences intracellular or extracellular expression of the factors of the present invention by yeast cells. [See, e.g., procedures

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described in published PCT application WO86/00639 and European patent application EPA 123,289].

A method for producing high levels of a BMP-9 protein of the invention in mammalian cells may involve the construction of cells containing multiple copies of the heterologous BMP-9 gene. The heterologous gene is linked to an amplifiable marker, e.g. the dihydrofolate reductase (DHFR) gene for which cells containing increased gene copies can be selected for propagation in increasing concentrations of methotrexate (MTX) according to the procedures of Kaufman and Sharp, J. Mol. Biol., 159:601-629 (1982). This approach can be employed with a number of different cell types.

For example, a plasmid containing a DNA sequence for a BMP-9 of the invention in operative association with other plasmid sequences enabling expression thereof and the DHFR expression plasmid pAdA26SV(A)3 [Kaufman and Sharp, Mol. Cell. Biol., 2:1304 (1982)] can be co-introduced into DHFR-deficient CHO cells, DUKX-BII, by various methods including calcium phosphate coprecipitation and transfection, electroporation or protoplast fusion. DHFR expressing transformants are selected for growth in alpha media with dialyzed fetal calf serum, and subsequently selected for amplification by growth in increasing concentrations of MTX (e.g. sequential steps in 0.02, 0.2, 1.0 and 5uM MTX) as described in Kaufman et al., Mol Cell Biol., Transformants are cloned, and biologically 5:1750 (1983). active BMP-9 expression is monitored by the Rosen-modified Sampath - Reddi rat bone formation assay described above in Example III. BMP-9 expression should increase with increasing levels of MTX resistance. BMP-9 polypeptides are characterized using standard techniques known in the art such as pulse labeling with [355] methionine or cysteine and polyacrylamide gel electrophoresis. Similar procedures can be followed to produce other related BMP-9 proteins.

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#### A. BMP-9 Vector Construction

In order to produce human BMP-9 proteins of the invention DNA sequences encoding the mature region of the human BMP-9 protein may be joined to DNA sequences encoding the propeptide region of the murine BMP-9 protein. This murine/human hybrid DNA sequence is inserted into an appropriate expression vector and introduced into mammalian cells or other preferred eukaryotic or prokaryotic hosts by conventional genetic engineering techniques. The construction of this murine/human BMP-9 containing expression plasmid is described below.

A derivative of the human BMP-9 sequence (SEQ ID NO:8) comprising the nucleotide sequence from nucleotide #105 to #470 is specifically amplified. The following oligonucleotides are utilized as primers to allow the amplification of nucleotides #105 to #470 of the human BMP-9 sequence (SEQ ID NO:8) from clone pGEM-111 described above.

# #3 ATCGGGCCCCTTTTAGCCAGGCGGAAAAGGAG

## #4 AGCGAATTCCCCGCAGGCAGATACTACCTG

This procedure generates the insertion of the nucleotide sequence ATCGGGCCCCT immediately preceding nucleotide #105 and the insertion of the nucleotide sequence GAATTCGCT immediately following nucleotide #470. The addition of these sequences results in the creation of an Apa I and EcoR I restriction endonuclease site at the respective ends of the specifically amplified DNA fragment. The resulting 374 bp Apa I/EcoR I fragment is subcloned into the plasmid vector pGEM-7Zf(+) (Promega catalog# p2251) which has been digested with Apa I and EcoR I. The resulting clone is designated phBMP9mex-1.

The following oligonucleotides are designed on the basis of murine BMP-9 sequences (SEQ ID NO:1) and are modified to facilitate the construction of the murine/human expression plasmid referred to above:

#5

GATTCCGTCGACCACCATGTCCCCTGGGGCCTGGTCTAGATGGATACACAGCTGTGGGGCC

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#6 CCACAGCTGTGTATCCATCTAGACCAGGCCCCAGGGGACATGGTGGTCGACG
These oligonucleotides contain complimentary sequences which
upon addition to each other facilitate the annealing (base
pairing) of the two individual sequences, resulting in the
formation of a double stranded synthetic DNA linker (designated
LINK-1) in a manner indicated below:

This DNA linker (LINK-1) contains recognition sequences of restriction endonucleases needed to facilitate subsequent manipulations required to construct the murine/human expression plasmid, as well as sequences required for maximal expression of heterologous sequences in mammalian cell expression systems. More specifically (referring to the sequence numbering of oligonucleotide #5/LINK-1): nucleotides #1-#11 recognition sequences for the restriction endonucleases BamH I and Sal I, nucleotides #11-#15 allow for maximal expression of heterologuos sequences in mammallian cell expression systems, nucleotides #16-#31 correspond to nucleotides #610-#625 of the murine BMP-9 sequence (SEQ ID NO:1), nucleotides #32-#33 are inserted to facilitate efficient restriction digestion of two adjacent restriction endonuclease sites (EcoO109 I and Xba I), nucleotides #34-#60 correspond to nucleotides #1515-#1541 of the murine BMP-9 sequence (SEQ ID NO:1) except that nucleotide #58 of synthetic oligonucloetide #5 is a G rather than the A which appears at position #1539 of SEQ ID NO:1 (This nucleotide conversion results in the creation of an Apa I restriction WO 93/00432 PCT/US92/05374

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endonuclease recognition sequence, without altering the amino acid sequence it is intended to encode, to facilitate further manipulations of the murine/human hybrid expression plasmid. LINK-1 (the double stranded product of the annealing of oligonucleotides #5 and #6) is subcloned into the plasmid vector pGEM-7Zf(+) which has been digested with the restriction endonucleases Apa I and BamH I. This results in a plasmid in which the sequences normally present between the Apa I and BamH I sites of the pGEM-7Zf(+) plasmid polylinker are replaced with the sequences of LINK-1 described above. The resulting plasmid clone is designated pBMP-9link.

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pBMP-9link is digested with the restriction endonucleases BamH I and Xba I resulting in the removal nucleotides #1-#34 of LINK-1 (refer to the numbering of oligo #5). Clone ML14a, which contains an insert comprising the sequence set forth in SEQ ID NO:1, is also digested with the restriction endonucleases BamH I and Xba I resulting in the removal of sequences comprising nucloetides #1-#1515 of SEQUENCE ID NO:1 (murine BMP-9). This BamH I/Xba I fragment of mouse BMP-9 is isolated from the remainder of the ML14a plasmid clone and subcloned into the BamH I/Xba I sites generated by the removal of the synthetic linker sequences described above. The resulting clone is designated p302.

The p302 clone is digested with the restriction 25 endonuclease EcoOlO9 I resulting in the excision of nucloetides corresponding to nucleotides #621-#1515 of the murine BMP-9

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sequence (SEQ ID NO:1) and nucleotides #35-#59 of LINK-1 (refer to numbering of oligonucleotide #5). It should be noted that the Apa I restriction site created in LINK-1 by the A to G conversion described above is a subset of the recognition sequence of EcoOlO9 I, therefore digestion of p302 with EcoOlO9 I cleaves at the Apa I site as well as the naturally occuring murine EcoO109 I (location #619-#625 of SEQ ID NO:1) resulting in the excision of a 920 bp EcoOlO9 I/EcoOlO9 I (Apa I) fragment comprising the sequences described above. This 920 EcoOlO9 I/EcoOlO9 I (Apa I) fragment is isolated from the remainder of the p302 plasmid clone and subcloned into clone pBMP-9link which has been similarly digested with EcoOlO9 I. It should be noted that the nucleotides GG (#32-#33 of oligonucleotide #5) originally designed to facilitate a more complete digestion of the two adjacent restriction sites EcoOlO9 I and Xba I of LINK-1, which is now a part of pBMP-9link (described above), results in the creation of Dcm methylation recognition sequence. The restriction nuclease EcoOlO9 I is sensitive to Dcm methylation and therefore of (nucleotides #25-#31 cleavage of this sequence oligonucleotide #5/LINK-1) by the restriction endonuclease EcoOlO9 I is prevented at this site. Therefore the plasmid clone pBMP-9link is cleaved at the Apa I site but not at the EcoOlO9 I site upon digestion with the restriction endonuclease EcoOlO9 I as described above, preventing the intended removal of the sequences between the EcoOlO9 I and Xba I site of LINK-1

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(#32-#55 defined by the numbering of oligonucleotide #5). This results in the insertion of the 920 bp EcoOlO9 I/Apa I fragment at the EcoOlO9 I (Apa I) site of pBMP-9link. The resulting clone is designated p318.

Clone p318 is digested with the restriction endonucleases Sal I and Apa I, resulting in the excision of sequences comprising nucleotides #6-#56 of LINK-1 (refer to oligo #5 for location), nucleotides #621-#1515 of murine BMP-9 (SEQ ID NO:1), and nucleotides #35-#60 of LINK-1 (refer to oligo #5 for location). The resulting 972 bp Sal I/Apa I fragment described above is isolated from the remainder of the p318 plasmid clone and will be utilized in subsequent manipulations.

The clone phBMP9mex-1 (described above), which contains DNA sequences which encode the entire mature region and portions of the propeptide of the human BMP-9 protein, is digested with the restriction endonucleases Apa I and EcoR I. This results in the excision of a 374 bp fragment comprising nucleotides #105-#470 of the human BMP-9 sequence (SEQ ID NO:8) and the additional nucleotides of oligonucleotide primers #3 and #4 which contain the recognition sequences for the restriction endonucleases Apa I and EcoR I. This 374 bp Apa I/EcoR I fragment is combined with the 972 bp Sal I/Apa I fragment from pl38 (isolation described above) and ligated to the mammalian cell expression plasmid pED6 (a derivative of pEMC2β1) which has been digested with Sal I and EcoR I. The resulting clone is designated p324.

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The clone ML14a (murine BMP-9) is digested with EcoO109 I and Xba I to generate a fragment comprising nucleotides #621-#1515 of SEQ ID NO:1.

The following oligonucleotides are synthesized on an automated DNA synthesizer and combined such that their complimentary sequences can base pair (anneal) with each other to generate a double stranded synthetic DNA linker designated LINK-2:

### **#7 TCGACCACCATGTCCCCTGG**

10 #8 GCCCCAGGGGACATGGTGG

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This double stranded synthetic DNA linker (LINK-2) anneals in such a way that it generates single stranded ends which are compatible to DNA fragments digested with Sal I (one end) or EcoOlO9 I (the other end) as indicated below:

#7 TCGACCACCATGTCCCCTGG
GGTGGTACAGGGGACCCCG #8:

This LINK-2 synthetic DNA linker is ligated to the 895 bp EcoOl09 I/Xba I fragment comprising nucleotides #621-#1515 of murine BMP-9 (SEQ ID NO:1) described above. This results in a 915 bp Sal I/Xba I fragment.

The clone p324 is digested with Sal I/Xba I to remove sequences comprising nucleotides #6-#56 of LINK-1 (refer to oligo #5 for location) and nucleotides #621-#1515 of murine BMP-9 (SEQ ID NO:1). The sequences comprising nucleotides #35-#60 of LINK-1 (refer to oligo #5 for location) and the sequences comprising the 374 bp Apa I/EcoR I fragment (human BMP-9 sequences) derived from phBMP9mex-1 remain attached to

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the pED6 backbone. The 915 bp Sal I/Xba I fragment comprising LINK-2 sequences and nucleotides #621-#1515 of murine BMP-9 (SEQ ID NO:1) is ligated into the p324 clone from which the Sal I to Xba I sequences described above have been removed.

The resulting plasmid is designated BMP9fusion and comprises LINK-2, nucleotides #621-#1551 of murine BMP-9 (SEQ ID NO:1), nucleotides #35-#59 of LINK-1 (refer to the numbering of oligonucleotide #5), and the 374 bp Apa I/EcoR I fragment (human BMP-9) derived from clone pBMP9mex-1 (described above) inserted between the Sal I and EcoR I sites of the mammalian cell expression vector pED6.

BMP9 fusion is transfected into CHO cells using standard techniques known to those having ordinary skill in the art to create stable cell lines capable of expressing human BMP-9 protein. The cell lines are cultured under suitable culture conditions and the BMP-9 protein is isolated and purified from the culture medium.

## EXAMPLE V

# 20 <u>Biological Activity of Expressed</u> BMP-9

To measure the biological activity of the expressed BMP-9 proteins obtained in Example IV above, the proteins are recovered from the cell culture and purified by isolating the BMP-9 proteins from other proteinaceous materials with which they are co-produced as well as from other contaminants. The purified protein may be assayed in accordance with the rat bone

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formation assay described in Example III.

Purification is carried out using standard techniques known to those skilled in the art. It is contemplated, as with other BMP proteins, that purification may include the use of Heparin sephanose.

Protein analysis is conducted using standard techniques such as SDS-PAGE acrylamide [U.K. Laemmli, Nature 227:680 (1970)] stained with silver [R.R. Oakley, et al. Anal. Biochem. 105:361 (1980)] and by immunoblot [H. Towbin, et al. Proc. Natl. Acad. Sci. USA 76:4350 (1979)]

The foregoing descriptions detail presently preferred embodiments of the present invention. Numerous modifications and variations in practice thereof are expected to occur to those skilled in the art upon consideration of these descriptions. Those modifications and variations are believed to be encompassed within the claims appended hereto.

# (1) GENERAL INFORMATION:

- (i) APPLICANT: Wozney, John M. Celeste, Anthony
- (ii) TITLE OF INVENTION: BMP-9 COMPOSITIONS
- (iii) NUMBER OF SEQUENCES: 9
- (iv) CORRESPONDENCE ADDRESS:
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  - (D) STATE: MA
    (E) COUNTRY: US
    (F) ZIP: 02140

  - (v) COMPUTER READABLE FORM:
    - (A) MEDIUM TYPE: Floppy disk
    - (B) COMPUTER: IBM PC compatible
    - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
    - (D) SOFTWARE: PatentIn Release #1.0, Version #1.25
- (vi) CURRENT APPLICATION DATA:
  - (A) APPLICATION NUMBER: US
  - (B) FILING DATE:
  - (C) CLASSIFICATION:
- (viii) ATTORNEY/AGENT INFORMATION:
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  - (B) REGISTRATION NUMBER: 32,245
  - (C) REFERENCE/DOCKET NUMBER: GI 5186A
  - (ix) TELECOMMUNICATION INFORMATION:
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    - (B) TELEFAX: (617) 876-5851
- (2) INFORMATION FOR SEQ ID NO:1:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 2447 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA to mRNA
  - (iii) HYPOTHETICAL: NO
    - (iv) ANTI-SENSE: NO
    - (vi) ORIGINAL SOURCE:
      - (A) ORGANISM: Mus musculus
      - (B) STRAIN: C57B46xCBA
      - (F) TISSUE TYPE: liver

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(vii) IMMEDIATE SOURCE:

(A) LIBRARY: Mouse liver cDNA (B) CLONE: ML14A

(viii) POSITION IN GENOME:

(C) UNITS: bp

(ix) FEATURE:

(A) NAME/KEY: mat\_peptide

(B) LOCATION: 1564..1893

(ix) FEATURE:

(A) NAME/KEY: CDS
(B) LOCATION: 610..1896

(ix) FEATURE:

(A) NAME/KEY: mRNA (B) LOCATION: 1..2447

(xi) SEQUENCE DESCRIPTION: SEQ ID NO:1:

CATTAATAAA TATTAAGTAT TGGAATTAGT GAAATTGGAG TTCCTTGTGG AAGGAAGTGG	60
GCAAGTGAGC TTTTTAGTTT GTGTCGGAAG CCTGTAATTA CGGCTCCAGC TCATAGTGGA	120
ATGGCTATAC TTAGATTTAT GGATAGTTGG GTAGTAGGTG TAAATGTATG TGGTAAAAGG	180
CCTAGGAGAT TTGTTGATCC AATAAATATG ATTAGGGAAA CAATTATTAG GGTTCATGTT	240
CGTCCTTTTG GTGTGGGAT TAGCATTATT TGTTTGATAA TAAGTTTAAC TAGTCAGTGT	300
TGGAAAGAAT GGAGACGGTT GTTGATTAGG CGTTTTGAGG ATGGGAATAG GATTGAAGGA	360
AATATAATGA TGGCTACAAC GATTGGGAAT CCTATTATTG TTGGGGTAAT GAATGAGGCA	420
AATAGATTTT CGTTCATTTT AATTCTCAAG GGGTTTTTAC TTTTATGTTT GTTAGTGATA	480
TTGGTGAGTA GGCCAAGGGT TAATAGTGTA ATTGAATTAT AGTGAAATCA TATTACTAGA	540
CCTGATGTTA GAAGGAGGC TGAAAAGGCT CCTTCCCTCC CAGGACAAAA CCGGAGCAGG	600
GCCACCCGG ATG TCC CCT GGG GCC TTC CGG GTG GCC CTG CTC CCG CTG  Met Ser Pro Gly Ala Phe Arg Val Ala Leu Leu Pro Leu  -318 -315 -310	648
TTC CTG CTG GTC TGT GTC ACA CAG CAG AAG CCG CTG CAG AAC TGG GAA  Phe Leu Leu Val Cys Val Thr Gln Gln Lys Pro Leu Gln Asn Trp Glu -305 -300 -295	696
CAA GCA TCC CCT GGG GAA AAT GCC CAC AGC TCC CTG GGA TTG TCT GGA Gln Ala Ser Pro Gly Glu Asn Ala His Ser Ser Leu Gly Leu Ser Gly -285 -280 -275	744
	792
GCT GGA GAG GAG GGT GTC TTT GAC CTG CAG ATG TTC CTG GAG AAC ATG Ala Gly Glu Gly Val Phe Asp Leu Gln Met Phe Leu Glu Asn Met -270 -265 -260	

# (2) INFORMATION FOR SEQ ID NO:9:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 151 amino acids
  - (B) TYPE: amino acid
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:9:
- \* Thr Arg Glu Cys Ser Arg Ser Cys Pro Arg Thr Ala Pro Gln Arg
  -41 -40 -35 -30
- Gln Val Arg Ala Val Thr Arg Arg Thr Arg Met Ala His Val Ala Ala -25 -15 -10
- Gly Ser Thr Leu Ala Arg Arg Lys Arg Ser Ala Gly Ala Gly Ser His
- Cys Gln Lys Thr Ser Leu Arg Val Asn Phe Glu Asp Ile Gly Trp Asp
- Ser Trp Ile Ile Ala Pro Lys Glu Tyr Glu Ala Tyr Glu Cys Lys Gly
- Gly Cys Phe Phe Pro Leu Ala Asp Asp Val Thr Pro Thr Lys His Ala 40 45 50 55
- Ile Val Gln Thr Leu Val His Leu Lys Phe Pro Thr Lys Val Gly Lys
  60 65 70
- Ala Cys Cys Val Pro Thr Lys Leu Ser Pro Ile Ser Val Leu Tyr Lys
  75 80 85
- Asp Asp Met Gly Val Pro Thr Leu Lys Tyr His Tyr Glu Gly Met Ser
- Val Ala Glu Cys Gly Cys Arg

(ix) FEATURE:	
(A) NAME/KEY: exon (B) LOCATION: 1470	
(ix) FEATURE: (A) NAME/KEY: CDS (B) LOCATION: 1456	•
(ix) FEATURE: (A) NAME/KEY: mat_peptide (B) LOCATION: 124453	
(ix) FEATURE: (A) NAME/KEY: mRNA (B) LOCATION: 1470	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:8:	
TGA ACA AGA GAG TGC TCA AGA AGC TGT CCA AGG ACG GCT CCA CAG AGG  * Thr Arg Glu Cys Ser Arg Ser Cys Pro Arg Thr Ala Pro Gln Arg  -41 -40 -35	48
CAG GTG AGA GCA GTC ACG AGG AGG ACA CGG ATG GCG CAC GTG GCT GCG Gln Val Arg Ala Val Thr Arg Arg Thr Arg Met Ala His Val Ala Ala -20 -15	96
GGG TCG ACT TTA GCC AGG CGG AAA AGG AGC GCC GGG GCT GGC AGC CAC Gly Ser Thr Leu Ala Arg Arg Lys Arg Ser Ala Gly Ala Gly Ser His	144
TGT CAA AAG ACC TCC CTG CGG GTA AAC TTC GAG GAC ATC GGC TGG GAC Cys Gln Lys Thr Ser Leu Arg Val Asn Phe Glu Asp Ile Gly Trp Asp 10 15 20	192
AGC TGG ATC ATT GCA CCC AAG GAG TAT GAA GCC TAC GAG TGT AAG GGC Ser Trp Ile Ile Ala Pro Lys Glu Tyr Glu Ala Tyr Glu Cys Lys Gly 25 30 35	240
GGC TGC TTC TTC CCC TTG GCT GAC GAT GTG ACG CCG ACG AAA CAC GCT Gly Cys Phe Phe Pro Leu Ala Asp Asp Val Thr Pro Thr Lys His Ala 40 45 50	288
ATC GTG CAG ACC CTG GTG CAT CTC AAG TTC CCC ACA AAG GTG GGC AAG Ile Val Gln Thr Leu Val His Leu Lys Phe Pro Thr Lys Val Gly Lys 60 65 70	336
GCC TGC TGT GTG CCC ACC AAA CTG AGC CCC ATC TCC GTC CTC TAC AAG Ala Cys Cys Val Pro Thr Lys Leu Ser Pro Ile Ser Val Leu Tyr Lys 75 80 85	364
GAT GAC ATG GGG GTG CCC ACC CTC AAG TAC CAT TAC GAG GGC ATG AGC ASP ASP Met Gly Val Pro Thr Leu Lys Tyr His Tyr Glu Gly Met Ser 90 95	432
GTG GCA GAG TGT GGG TGC AGG TAGTATCTGC CTGCGGG Val Ala Glu Cys Gly Cys Arg 105	470

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#### CATGGGCAGC TCGAG

(2)	INFORMATION	FOR	SEO	ID	NO:6:
-----	-------------	-----	-----	----	-------

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 34 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:6: CTGCAGGCGA GCCTGAATTC CTCGAGCCAT CATG

34

- (2) INFORMATION FOR SEQ ID NO:7:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 68 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double

    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: cDNA to mRNA
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO:7:

CGAGGTTAAA AAACGTCTAG GCCCCCGAA CCACGGGGAC GTGGTTTTCC TTTGAAAAAC 60 ACGATTGC 68

- (2) INFORMATION FOR SEQ ID NO:8:
  - (i) SEQUENCE CHARACTERISTICS:
    - (A) LENGTH: 470 base pairs
    - (B) TYPE: nucleic acid
    - (C) STRANDEDNESS: double
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: DNA (genomic)
  - (iii) HYPOTHETICAL: NO
    - (v) FRAGMENT TYPE: C-terminal
    - (vi) ORIGINAL SOURCE:
      - (A) ORGANISM: Homo sapiens
      - (H) CELL LINE: W138 (genomic DNA)
  - (vii) IMMEDIATE SOURCE:
    - (A) LIBRARY: human genomic library
    - (B) CLONE: lambda 111-1
  - (viii) POSITION IN GENOME:
    - (C) UNITS: bp

Gly His Leu Ile Thr Arg Leu Leu Asp Thr Arg Leu Val His His Asn -95

Val Thr Arg Trp Glu Thr Phe Asp Val Ser Pro Ala Val Leu Arg Trp -70

Arg Ile Asn Ile Tyr Glu Val Met Lys Pro Pro Ala Glu Val Val Pro

Thr Arg Glu Lys Gln Pro Asn Tyr Gly Leu Ala Ile Glu Val Thr His

Leu His Gln Thr Arg Thr His Gln Gly Gln His Val Arg Ile Ser Arg

Ser Leu Pro Gln Gly Ser Gly Asn Trp Ala Gln Leu Arg Pro Leu Leu
-35 -30 -25

Val Thr Phe Gly His Asp Gly Arg Gly His Ala Leu Thr Arg Arg Arg -20 -15 -10 -5

Arg Ala Lys Arg Ser Pro Lys His His Ser Gln Arg Ala Arg Lys Lys
1 10

Asn Lys Asn Cys Arg Arg His Ser Leu Tyr Val Asp Phe Ser Asp Val 15 20 25

Gly Trp Asn Asp Trp Ile Val Ala Pro Pro Gly Tyr Gln Ala Phe Tyr . 30 35 40

Cys His Gly Asp Cys Pro Phe Pro Leu Ala Asp His Leu Asn Ser Thr 45 50 55 60

Asn His Ala Ile Val Gln Thr Leu Val Asn Ser Val Asn Ser Ser Ile
65 70 75

Pro Lys Ala Cys Cys Val Pro Thr Glu Leu Ser Ala Ile Ser Met Leu 80 85 90

Tyr Leu Asp Glu Tyr Asp Lys Val Val Leu Lys Asn Tyr Gln Glu Met 95 100 105

Val Val Glu Gly Cys Gly Cys Arg

### (2) INFORMATION FOR SEQ ID NO:5:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 15 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:5:

TGT GGG TGC CGC TGAGATCAGG CAGTCCTTGA GGATAGACAG ATATACACAC 1 Cys Gly Cys Arg 115	.666
CACACACA CACCACATAC ACCACACACA CACGTTCCCA TCCACTCACC CACACACTAC 1	726
ACAGACTGCT TCCTTATAGC TGGACTTTTA TTTAAAAAAA AAAAAAAAA AATGGAAAAA 1	.786
ATCCCTAAAC ATTCACCTTG ACCTTATTTA TGACTTTACG TGCAAATGTT TTGACCATAT	.846
TGATCATATA TTTTGACAAA ATATATTTAT AACTACGTAT TAAAAGAAAA AAATAAAATG	906
AGTCATTATT TTAAAAAAA AAAAAAACT CTAGAGTCGA CGGAATTC	L954
(2) INFORMATION FOR SEQ ID NO:4:  (i) SEQUENCE CHARACTERISTICS:  (A) LENGTH: 408 amino acids  (B) TYPE: amino acid  (D) TOPOLOGY: linear	
(ii) MOLECULE TYPE: protein	
(xi) SEQUENCE DESCRIPTION: SEQ ID NO:4:	
Met Ile Pro Gly Asn Arg Met Leu Met Val Val Leu Leu Cys Gln Val -292 -280 -280	
Leu Leu Gly Gly Ala Ser His Ala Ser Leu Ile Pro Glu Thr Gly Lys -275 -270 -265	
Lys Lys Val Ala Glu Ile Gln Gly His Ala Gly Gly Arg Arg Ser Gly -260 -255 -250 -245	
Gln Ser His Glu Leu Leu Arg Asp Phe Glu Ala Thr Leu Leu Gln Met -240 -235 -230	
Phe Gly Leu Arg Arg Pro Gln Pro Ser Lys Ser Ala Val Ile Pro -225 -220 -215	
Asp Tyr Met Arg Asp Leu Tyr Arg Leu Gln Ser Gly Glu Glu Glu Glu -200 -200	
Glu Gln Ile His Ser Thr Gly Leu Glu Tyr Pro Glu Arg Pro Ala Ser -195 -190 -185	
Arg Ala Asn Thr Val Arg Ser Phe His His Glu Glu His Leu Glu Asn -180 -175 -170 -165	
Ile Pro Gly Thr Ser Glu Asn Ser Ala Phe Arg Phe Leu Phe Asn Leu -160 -155 -150	

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Ser Ser Ile Pro Glu Asn Glu Val Ile Ser Ser Ala Glu Leu Arg Leu

Phe Arg Glu Gln Val Asp Gln Gly Pro Asp Trp Glu Arg Gly Phe His -130 -125 -120

-140

				Pro					Gly			CGT Arg		Asn		942
			Met					Glu				GGG Gly -100	His			990
												GTG Vaļ				1038
												ACC Thr				1086
CAG Gln	CCA Pro	AAC Asn	TAT Tyr	GGG Gly -60	CTA Leu	GCC Ala	ATT Ile	GAG Glu	GTG Val -55	ACT Thr	CAC His	CTC Leu	CAT His	CAG Gln -50	ACT Thr	1134
												TCG Ser				1182
												GTĆ Val -20				1230
												AGG Arg				1278
												AAT Asn				1326
												GGC Gly				1374
												TGC Cys 45				1422
												AAC Asn				1470
						Ser						CCC Pro				1518
					Leu							TAC Tyr				1566
				Val					Gln			GTA Val				1614

(B) LOCATION: 9..1934

#### (xi) SEQUENCE DESCRIPTION: SEQ ID NO:3:

CTCTAGAGGG CAGAGGAGGA GGGAGGGAGG GAAGGAGCGC GGAGCCCGGC CC	EGGAAGCTA 60
GGTGAGTGTG GCATCCGAGC TGAGGGACGC GAGCCTGAGA CGCCGCTGCT GC	TCCGGCTG 120
AGTATCTAGC TTGTCTCCCC GATGGGATTC CCGTCCAAGC TATCTCGAGC CT	GCAGCGCC 180
ACAGTCCCCG GCCCTCGCCC AGGTTCACTG CAACCGTTCA GAGGTCCCCA GG	AGCTGCTG 240
CTGGCGAGCC CGCTACTGCA GGGACCTATG GAGCCATTCC GTAGTGCCAT CO	CCGAGCAAC 300
GCACTGCTGC AGCTTCCCTG AGCCTTTCCA GCAAGTTTGT TCAAGATTGG CT	GTCAAGAA 360
TCATGGACTG TTATTATATG CCTTGTTTTC TGTCAAGACA CC ATG ATT CC Met Ile Pr -292 -2	
AAC CGA ATG CTG ATG GTC GTT TTA TTA TGC CAA GTC CTG CTA CAS ASN Arg Met Leu Met Val Val Leu Leu Cys Gln Val Leu Cys Gln Val Leu Cys Gl	
GCG AGC CAT GCT AGT TTG ATA CCT GAG ACG GGG AAG AAA AAA CAAA AAA CAAA Ser His Ala Ser Leu Ile Pro Glu Thr Gly Lys Lys Lys Caa Caa Caa Caa Caa Caa Caa Caa Caa Ca	
GAG ATT CAG GGC CAC GCG GGA GGA CGC CGC TCA GGG CAG AGC CGC GLU Ile Gln Gly His Ala Gly Gly Arg Arg Ser Gly Gln Ser 1 -255 -250 -245	
CTC CTG CGG GAC TTC GAG GCG ACA CTT CTG CAG ATG TTT GGG (Leu Leu Arg Asp Phe Glu Ala Thr Leu Leu Gln Met Phe Gly : -235 -230	
CGC CGC CCG CAG CCT AGC AAG AGT GCC GTC ATT CCG GAC TAC Arg Arg Pro Gln Pro Ser Lys Ser Ala Val Ile Pro Asp Tyr : -220 -215	
GAT CTT TAC CGG CTT CAG TCT GGG GAG GAG GAG GAA GAG CAG Asp Leu Tyr Arg Leu Gln Ser Gly Glu Glu Glu Glu Gln -205 -200 -195	Ile His
AGC ACT GGT CTT GAG TAT CCT GAG CGC CCG GCC AGC CGG GCC Ser Thr Gly Leu Glu Tyr Pro Glu Arg Pro Ala Ser Arg Ala -190 -185 -180	AAC ACC 750 Asn Thr
GTG AGG AGC TTC CAC CAC GAA GAA CAT CTG GAG AAC ATC CCA Val Arg Ser Phe His His Glu Glu His Leu Glu Asn Ile Pro -175 -170 -165	
AGT GAA AAC TCT GCT TTT CGT TTC CTC TTT AAC CTC AGC AGC Ser Glu Asn Ser Ala Phe Arg Phe Leu Phe Asn Leu Ser Ser -160 -155 -150	ATC CCT 846 Ile Pro -145
GAG AAC GAG GTG ATC TCC TCT GCA GAG CTT CGG CTC TTC CGG Glu Asn Glu Val Ile Ser Ser Ala Glu Leu Arg Leu Phe Arg -140 -135	GAG CAG 894 Glu Gln -130

-10

Gly Ala Ser Ser His Cys Gln Lys Thr Ser Leu Arg Val Asn Phe Glu
5 10 15

Asp Ile Gly Trp Asp Ser Trp Ile Ile Ala Pro Lys Glu Tyr Asp Ala 20 25 30

Tyr Glu Cys Lys Gly Gly Cys Phe Phe Pro Leu Ala Asp Asp Val Thr 35 40 45 50

Pro Thr Lys His Ala Ile Val Gln Thr Leu Val His Leu Glu Phe Pro 55 60 65

Thr Lys Val Gly Lys Ala Cys Cys Val Pro Thr Lys Leu Ser Pro Ile 70 75 80

Ser Ile Leu Tyr Lys Asp Asp Met Gly Val Pro Thr Leu Lys Tyr His
85 90 95

Tyr Glu Gly Met Ser Val Ala Glu Cys Gly Cys Arg

#### (2) INFORMATION FOR SEQ ID NO:3:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1954 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA to mRNA
- (iii) HYPOTHETICAL: NO
- (iv) ANTI-SENSE: NO
- (vi) ORIGINAL SOURCE:
  - (A) ORGANISM: Homo sapiens
  - (G) CELL TYPE: Osteosarcoma Cell Line
  - (H) CELL LINE: U-20S
- (vii) IMMEDIATE SOURCE:
  - (A) LIBRARY: U2OS cDNA in Lambda gt10
  - (B) CLONE: Lambda U205-3
- (viii) POSITION IN GENOME:
  - (C) UNITS: bp
  - (ix) FEATURE:
    - (A) NAME/KEY: CDS
    - (B) LOCATION: 403..1629
  - (ix) FEATURE:
    - (A) NAME/KEY: mat\_peptide
    - (B) LOCATION: 1279..1626
  - (ix) FEATURE:
    - (A) NAME/KEY: mRNA

- (xi) SEQUENCE DESCRIPTION: SEQ ID NO:2:
- Met Ser Pro Gly Ala Phe Arg Val Ala Leu Leu Pro Leu Phe Leu Leu -318 -315 -305
- Val Cys Val Thr Gln Gln Lys Pro Leu Gln Asn Trp Glu Gln Ala Ser
  -300 -295 -290
- Pro Gly Glu Asn Ala His Ser Ser Leu Gly Leu Ser Gly Ala Gly Glu
  -285 -280 -275
- Glu Gly Val Phe Asp Leu Gln Met Phe Leu Glu Asn Met Lys Val Asp
  -270 -265 -260 -255
- Phe Leu Arg Ser Leu Asn Leu Ser Gly Ile Pro Ser Gln Asp Lys Thr
  -250 -245 -240
- Arg Ala Glu Pro Pro Gln Tyr Met Ile Asp Leu Tyr Asn Arg Tyr Thr
  -235 -230 -225
- Thr Asp Lys Ser Ser Thr Pro Ala Ser Asn Ile Val Arg Ser Phe Ser -220 -215 -210
- Val Glu Asp Ala Ile Ser Thr Ala Ala Thr Glu Asp Phe Pro Phe Gln -205 -200 -195
- Lys His Ile Leu Ile Phe Asn Ile Ser Ile Pro Arg His Glu Gln Ile
  -190 -185 -180 -175
- Thr Arg Ala Glu Leu Arg Leu Tyr Val Ser Cys Gln Asn Asp Val Asp
  -170 -165 -160
- Ser Thr His Gly Leu Glu Gly Ser Met Val Val Tyr Asp Val Leu Glu
  -155 -150 -145
- Asp Ser Glu Thr Trp Asp Gln Ala Thr Gly Thr Lys Thr Phe Leu Val -140 -135 -130
- Ser Gln Asp Ile Arg Asp Glu Gly Trp Glu Thr Leu Glu Val Ser Ser -125 -120 -115
- Ala Val Lys Arg Trp Val Arg Ala Asp Ser Thr Thr Asn Lys Asn Lys -110 -105 -100 -95
- Leu Glu Val Thr Val Gln Ser His Arg Glu Ser Cys Asp Thr Leu Asp
  -90 -85
- Ile Ser Val Pro Pro Gly Ser Lys Asn Leu Pro Phe Phe Val Val Phe -75 -70 -65
- Ser Asn Asp Arg Ser Asn Gly Thr Lys Glu Thr Arg Leu Glu Leu Lys
  -60 -55
- Glu Met Ile Gly His Glu Gln Glu Thr Met Leu Val Lys Thr Ala Lys
  -45 -40 -35
- Asn Ala Tyr Gln Val Ala Gly Glu Ser Gln Glu Glu Glu Gly Leu Asp
  -30 -25 -20 -15
- Gly Tyr Thr Ala Val Gly Pro Leu Leu Ala Arg Arg Lys Arg Ser Thr

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	-15					-10					-5					
AGG AGC Arg Ser 1	ACC Thr	GGA Gly	GCC Ala	AGC Ser 5	AGC Ser	CAC His	TGC Cys	CAG Gln	AAG Lys 10	ACT Thr	TCT Ser	CTC Leu	AGG Arg	GTG Val 15	1608	1
AAC TTT Asn Phe	GAG Glu	GAC Asp	ATC Ile 20	GGC Gly	TGG Trp	GAC Asp	AGC Ser	TGG Trp 25	ATC Ile	ATT Ile	GCA Ala	CCC Pro	AAG Lys 30	GAA Glu	1656	i
TAT GAC Tyr Asp	GCC Ala	TAT Tyr 35	GAG Glu	TGT Cys	AAA Lys	GGG Gly	GGT Gly 40	TGC Cys	TTC Phe	TTC Phe	cca Pro	TTG Leu 45	GCT Ala	GAT Asp	1704	ı
GAC GTG Asp Val	ACA Thr 50	CCC Pro	ACC Thr	AAA Lys	CAT His	GCC Ala 55	ATC Ile	GTG Val	CAG Gln	ACC Thr	CTG Leu 60	GTG Val	CAT His	CTC Leu	1752	!
GAG TTC Glu Phe 65	CCC Pro	ACA Thr	AAG Lys	GTG Val	GGC Gly 70	AAA Lys	GCC Ala	TGC Cys	TGC Cys	GTT Val 75	ccc Pro	ACC Thr	AAA Lys	CTG Leu	1800	)
AGT CCC Ser Pro 80	ATC Ile	TCC Ser	ATC Ile	CTC Leu 85	TAC Tyr	AAG Lys	GAT Asp	GAC Asp	ATG Met 90	GGG	GTG Val	CCA Pro	ACC Thr	CTC Leu 95	1848	!
AAG TAC Lys Tyr	CAC His	Tyr	GAG Glu 100	GGG Gly	ATG Meț	AGT Ser	GTG Val	GCT Ala 105	GAG Glu	TGT Cys	GGG Gly	TGT Cys	AGG Arg 110	TAGTCCCI	:GC	1903
AGCCACC	CAG (	GGTG	GGA!	ra ci	AGGA	CATG	AA C	GAGG:	TTCT	GGT	ACGG:	rcc '	TGCAT	CCTCC	1963	3
TGCGCATO	GT 1	ATGC	CTAAC	T T	GATC	AGAA	A CC	ATCC:	ITGA	GAA	GAAAI	AGG .	AGTT	AGTTGC	2023	3
CCTTCTTC	GTG !	TCTG	GTGG	T C	CCTC	rgct	AA E	GTGA	CAAT	GAC	rggg	GTA '	TGCG	GCCTG	2083	3
TGGGCAG	AGC 1	AGGA	GACC	CT G	GAAG	GTT	A GT	GGGT	AGAA	AGA:	rgtc	AAA .	AAGG	AAGCTG	2143	3
TGGGTAG	ATG 1	ACCT	GCAC:	re c	AGTG!	ATTA	AA(	GTCC	AGCC	TTA	CCTG:	rga ·	GAGA	SCTCCT	2203	3
GGCATCT	AAG 1	AGAA(	CTCT	C T	rccT(	CATC	A TC	CCCA	CCGA	CTT	GTTC:	TTC	CTTG	GAGTG	2263	3
TGTCCTC	AGG (	GAGA	ACAG	CA T	rgcT(	STTC	C TG	rgcc:	rcaa	GCT	CCA	SCŤ (	GACT	CTCCTG	2323	3
TGGCTCAT	PAG (	GACT	GAAT	GG G	GTGA	GGAA	G AG	CCTG	ATGC	CCT	CTGG	CAA	TCAG	AGCCCG	2383	3
AAGGACT	rca i	AAAC	ATCT	G A	CAAC	rctc2	A TT	GACT	GATG	CTC	CAAC	ATA .	ATTT	AAAATI	2443	3
AGAG															2447	,

### (2) INFORMATION FOR SEQ ID NO:2:

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 428 amino acids
    (B) TYPE: amino acid
    (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein

-250 -255 -245 GAC AAA ACC AGA GCG GAG CCA CCC CAG TAC ATG ATC GAC TTG TAC AAC 888 Asp Lys Thr Arg Ala Glu Pro Pro Gln Tyr Met Ile Asp Leu Tyr Asn -235 -240 AGA TAC ACA ACG GAC AAA TCG TCT ACG CCT GCC TCC AAC ATC GTG CGG 936 Arg Tyr Thr Thr Asp Lys Ser Ser Thr Pro Ala Ser Asn Ile Val Arg -225 -220 AGC TTC AGC GTG GAA GAT GCT ATA TCG ACA GCT GCC ACG GAG GAC TTC 984 Ser Phe Ser Val Glu Asp Ala Ile Ser Thr Ala Ala Thr Glu Asp Phe -205 CCC TTT CAG AAG CAC ATC CTG ATC TTC AAC ATC TCC ATC CCG AGG CAC 1032 Pro Phe Gln Lys His Ile Leu Ile Phe Asn Ile Ser Ile Pro Arg His -185 -190 GAG CAG ATC ACC AGG GCT GAG CTC CGA CTC TAT GTC TCC TGC CAA AAT 1080 Glu Gln Ile Thr Arg Ala Glu Leu Arg Leu Tyr Val Ser Cys Gln Asn -175 -170GAT GTG GAC TCC ACT CAT GGG CTG GAA GGA AGC ATG GTC GTT TAT GAT 1128 Asp Val Asp Ser Thr His Gly Leu Glu Gly Ser Met Val Val Tyr Asp -155 -160 1176 GTT CTG GAG GAC AGT GAG ACT TGG GAC CAG GCC ACG GGG ACC AAG ACC Val Leu Glu Asp Ser Glu Thr Trp Asp Gln Ala Thr Gly Thr Lys Thr -130 -145 TTC TTG GTA TCC CAG GAC ATT CGG GAC GAA GGA TGG GAG ACT TTA GAA 1224 Phe Leu Val Ser Gln Asp Ile Arg Asp Glu Gly Trp Glu Thr Leu Glu -120 -125 GTA TCG AGT GCC GTG AAG CGG TGG GTC AGG GCA GAC TCC ACA ACA AAC 1272 Val Ser Ser Ala Val Lys Arg Trp Val Arg Ala Asp Ser Thr Thr Asn -110 AAA AAT AAG CTC GAG GTG ACA GTG CAG AGC CAC AGG GAG AGC TGT GAC 1320 Lys Asn Lys Leu Glu Val Thr Val Gln Ser His Arg Glu Ser Cys Asp ACA CTG GAC ATC AGT GTC CCT CCA GGT TCC AAA AAC CTG CCC TTC TTT 1368 Thr Leu Asp Ile Ser Val Pro Pro Gly Ser Lys Asn Leu Pro Phe Phe GTT GTC TTC TCC AAT GAC CGC AGC AAT GGG ACC AAG GAG ACC AGA CTG 1416 Val Val Phe Ser Asn Asp Arg Ser Asn Gly Thr Lys Glu Thr Arg Leu -60 1464 GAG CTG AAG GAG ATG ATC GGC CAT GAG CAG GAG ACC ATG CTT GTG AAG Glu Leu Lys Glu Met Ile Gly His Glu Gln Glu Thr Met Leu Val Lys 1512 ACA GCC AAA AAT GCT TAC CAG GTG GCA GGT GAG AGC CAA GAG GAG Thr Ala Lys Asn Ala Tyr Gln Val Ala Gly Glu Ser Gln Glu Glu GGT CTA GAT GGA TAC ACA GCT GTG GGA CCA CTT TTA GCT AGA AGG AAG 1560 Gly Leu Asp Gly Tyr Thr Ala Val Gly Pro Leu Leu Ala Arg Arg Lys

What is claimed is:

- A BMP-9 polypeptide comprising the amino acid sequence from amino acid #8 - 110 as set forth in FIG. 3 (SEQ ID NO: 9).
- 2. A BMP-9 polypeptide comprising the amino acid sequence from amino acid #1 110 as set forth in FIG. 3 (SEQ ID NO: 9).
- 3. A BMP-9 polypeptide of claim 1 wherein said polypeptide is a dimer wherein each subunit comprises at least the amino acid sequence from amino acid #8 110 of FIG. 3 (SEQ ID NO: 9).
- 4. A BMP-9 polypeptide of claim 2 wherein said polypeptide is a dimer wherein each subunit comprises at least the amino acid sequence from amino acid #1-110 of FIG. 3. (SEQ ID NO: 9).
- 5. A purified BMP-9 protein produced by the steps of
- (a) culturing a cell transformed with a cDNA comprising the nucleotide sequence from nucleotide #124 to #453 as shown in FIG. 3 (SEQ ID NO: 8); and
- (b) recovering and purifying from said culture medium a protein comprising the amino acid sequence from amino acid #1 to amino acid #110 as shown in FIG. 3 (SEQ ID NO: 9).
- 6. A purified BMP-9 protein produced by the steps of
  - (a) culturing a cell transformed with a cDNA comprising

the nucleotide sequence from nucleotide #124 to #453 as shown in FIG. 3 (SEQ ID NO: 8); and

- (b) recovering form said culture medium a protein comprising an amino acid sequence from amino acid #8 to amino acid #110 as shown in Figure 3 (SEQ ID NO: 9).
- 7. A BMP-9 protein characterized by the ability to induce the formation of cartilage and/or bone.
- 8. A DNA sequence encoding a BMP-9 protein.
- 9. The DNA sequence of claim 8 wherein said DNA comprises
  - (a) nucleotide 124 to 453 (SEQ ID NO: 8); and
- (b) sequences which hybridize thereto under stringent hybridization conditions and exhibit the ability to form cartilage and/or bone.
- 10. The DNA sequence of claim 8 wherein said DNA comprises
  - (a) nucleotide 145 to 453 (SEQ ID NO: 8); and
- (b) sequences which hybridize thereto under stringent hybridization conditions and exhibit the ability to form cartilage and/or bone.
- 11. A host cell transformed with a DNA sequence encoding BMP-8.

- 12. A method for producing a purified BMP-9 protein said method comprising the steps of
- (a) culturing a cell transformed with a cDNA comprising the nucleotide sequence encoding a BMP-9 protein; and
- (b) recovering and purifying said BMP-9 protein from the culture medium.
- 13. A pharmaceutical composition comprising an effective amount of a BMP-9 protein in admixture with a pharmaceutically acceptable vehicle.
- 14. A composition of claim 13 further comprising a matrix for supporting said composition and providing a surface for bone and/or cartilage growth.
- 15. The composition of claim 14 wherein said matrix comprises a material selected from the group consisting of hydroxyapatite, collagen, polylactic acid and tricalcium phosphate.
- 16. A method for inducing bone and/or cartilage formation in a patient in need of same comprising administering to said patient an effective amount of the composition of claim 13.
- 17. A pharmaceutical composition for wound healing and tissue repair said composition comprising an effective amount of the

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protein of a BMP-9 protein in a pharmaceutically acceptable vehicle.

18. A method for treating wounds and/or tissue repair in a patient in need of same comprising administering to said patient an effective amount of the composition of claim 17.

# Figure 1A

		.0		20			30			40		50			60		70	)
CATTA	ATAA	A TA	TTAA	GTAT	TGG.	AATT.	AGT	GAAAT	rtgg!	AG T	TCCTI	GTGG	AAG	GAAG	TGG (	GCAA	STGAGO	2
		10		0.0			100			• •		• • •						
لعزمليني			יכייירכ	90 30 AG			100 דירים	caaci		10 30 T	ር እጥ አር	120 maga		COTTA	130	mm a c :	14 ATTTAT	.0
	15		0100	160			170			80	CALAC	190		GCIN	200	IIAGI	21 21	_
GGATA	GTTC	G GT	AGTA					TGGT			CTAGG			TTGA		ATAA	YTATAA	.u 3
	22	20		230	1		240		2	50		260	כ		270		28	0
ATTAG			ATTA					CGTC			TGTGI			CATT		TGTT:	IGATA	A
m > > < m	29		CEC.	300			310			20		330			340		35	
TAAGI	3 <i>6</i>		GTCA	GTGT 370			AAT ( 380	GGAGA		17. G	TTGAT	TAGG. 400		TITG		ATGG	SAATAC	
GATTO			מיתת					GAጥጥ(			רייועמיים			ссст	410	ሮኔ ኔጥ	42 GAGGC	
	43			440			450			60	~	470		0001	480	0.11.1	37.66C2	
AATAG	TTTA	T CG	TTCA	TTTT	AAT	TCTC	AAG	GGGT:	PTTT	AC T	TTTAT			AGTG		TTGG:	TGAGT	
	50	00		510	)		520		5	30		54	0		550		56	50
GGCCA	AGGG	AT T	ATAG			GAAT			TAAA					GATG			GAGGG	2
	5	70		58	30		59	0		600	1	60	-		6:	18		
тсаз	AAGO	- COTE (	الماليات	-C-T-C	יר כז	CCAC	מממי	N CC	20300	יא ככ	GCC		יר א <sup>י</sup>	<u> </u>	<del>-</del> -	<del></del>	<del></del>	
10.0	<i></i>				.c c,	NOON		n CC	JONG	JAGG	GCC	1000	M	S	P		<b>.</b>	
												_	••		•	·		
	627			636			645			654			663			672		
	<del></del>																	
. GCC A	TTC F	CGG R	GTG V	GCC		CTC					CTG					CAG		
A	r	K	V	A	L	L	P	L	F	L	L	V	С	V	T	Q	Q	
	681			690			699			708	:		717			726		
AAG	CCG	CTG	CAG	AAC	TGG	GAA	CAA	GCA	TCC	CCT	GGG	GAA	$\overline{\mathtt{AAT}}$	GCC	CAC	AGC	TCC	
K	P	L	Q	N	W	E	Q	A	S	P	G	E	N	A	H	S	S	
	735			- 4 4										•				
	/35			744			753			762			771			780		
CTG	GGA	TTG	TCT	GGA	GCT	GGA	GAG	GAG	GGT	GTC	TTT	GAC	CTC	CAG	<del>M</del> TC	TTC	<del>cre</del>	
L	G	L	S	G	A	G	E	E	G	V	F	D	L	0	M	F	L	
						•	_	_	_	•	-	-	_	•	••	-	_	
	789			798			807			816	;		825			834		
<u> </u>	770	<u> </u>	776	<del></del>	<del></del>	<del></del>		===	<del></del>						\ <u></u>		===	
E E	AAC N	ATG M	_								AAC						TCC	
	F4	11	Λ.		1.7	r		R		1.	FN:	4.			3	-		

## Figure 1B

	843			852			861			870			879			888	
CAG Q	GAC D	AAA K	ACC T	AGA R	GCG A	GAG E	CCA P	CCC P	CAG Q	TAC Y	ATG M	ATC I	GAC D	TTG L	TAC Y	AAC N	ĀGĀ R
	897			906			915			924			933			942	
TAC Y	ACA T	ACG T	GAC D	AAA K	TCG S	TCT S	ACG T	CCT P	GCC A	TCC S	AAC N	ATC I	GTG V	CGG R	AGC S	TTC F	AGC S
	951			960			969			978			987			996	
GTG V	GAA E	GAT D	GCT A	ATA I	TCG S	ACA T	GCT A	GCC A	ACG T	GAG E	GAC D	TTC F	CCC P	TTT F	CAG Q	AAG K	CAC H
:	1005		:	1014		:	1023		;	1032			1041		:	1050	
ATC I	CTG L	ATC I	TTC F	AAC N	ATC I	TCC S	ATC I	CCG P	AGG R	CAC H	GAG E	CAG Q	ATC I	ACC T	AGG R	GCT A	GAG E
;	1059		:	1068		:	1077		:	1086			1095		:	1104	
CTC L	CGA R	CTC L	$\overline{\underline{\mathtt{TAT}}}_{\underline{Y}}$	GTC V	TCC S	TGC C	CAA Q	AAT N	GAT D	GTG V	GAC D	TCC S	ACT T	CAT H	GGG G	CTG L	GAA E
																	_
:	1113		:	1122		:	1131		;	1140			1149		-	1158	_
		ATG M			TAT Y			CTG L		GAC	AGT S			TGG W		CAG Q	
GGA G.	AGC	ATG M	GTC V	GTT	<del>TAT</del> Y	GAT D	GTT	CTG L	GAG E	GAC	ĀGT S	GAG E	ACT		GAC D	CAG	GCC
GGA G	AGC S	M	GTC V	GTT V	Y	GAT D	GTT V	L	GAG E CAG	GAC D 1194	s	GAG E	ACT T	W	GAC D	CAG Q	GCC A
GGA G ACG	AGC S 1167 GGG	M ACC	GTC V :	GTT V 1176 ACC	Y TTC	GAT D TTG L	GTT V 1185 GTA	L TCC	GAG E CAG	GAC D 1194 GAC	S ATT	GAG E CGG R	ACT T 1203 GAC	W GAA	GAC D GGA G	CAG Q 1212 TGG	GCC A
GGA G ACG T	AGC S 1167 GGG G	M ACC T	GTC V AAG K	GTT V 1176 ACC T	TTC F	GAT D TTG L	GTT V 1185 GTA V	TCC S	GAG E CAG Q	GAC D 1194 GAC D	S ATT I	GAG E CGG R	ACT T 1203 GAC D	GAA E	GAC D	CAG Q 1212 TGG W	GCC A GAG E
GGA G ACG T ACT	AGC S 1167 GGG G 1221	ACC T	GTC V AAG K	GTT V 1176 ACC T 1230	TTC F	GAT D TTG L	GTT V 1185 GTA V 1239 GTG	TCC S	GAG E . CAG Q	GAC D 1194 GAC D 1248	S ATT I	GAG E CGG R AGG R	ACT T 1203 GAC D 1257 GCA	GAA E	GAC D GGA G	CAG Q 1212 TGG W 1266	GCC A GAG E
GGA G ACG T ACT	AGC S 1167 GGG G 1221 TTA L	ACC T GAA E	GTC V AAG K GTA V	GTT V 1176 ACC T 1230 TCG S	TTC F	GAT D TTG L	GTT V 1185 GTA V 1239 GTG V	TCC S AAG K	GAG E CAG Q	GAC D 1194 GAC D 1248 TGG W	ATT I GTC V	GAG E CGG R AGG R	ACT T 1203 GAC D 1257 GCA A	GAA E GAC D	GAC D GGA G	CAG Q 1212 TGG W 1266 ACA	GCC A GAG E
GGA G ACG T ACT T	AGC S 1167 GGG G 1221 TTA L 1275 AAA	ACC T  GAA E	GTC V AAG K GTA V	GTT V 1176 ACC T 1230 TCG S 1284 CTC	TTC F	GAT D TTG L GCC A	GTT V 1185 GTA V 1239 GTG V 1293 ACA	TCC S AAG K	GAG E . CAG Q	GAC D 1194 GAC D 1248 TGG W 1302	S ATT I GTC V	GAG E CGG R AGG R	ACT T 1203 GAC D 1257 GCA A	GAA E GAC D	GAC D GGA G TCC S	CAG Q 1212 TGG W 1266 ACA T	GCC A GAG E

# Figure 1C

	1383			1392			1401			1410			1419			1428	
TCC S	AAT N	GAC D	CGC R	AGC S	AAT N	GGG G	ACC T	AAG K	GAG E	ACC T	AGA R	CTG L	GAG E	CTG L	AAG K	GAG E	ĀTG M
	1437			1446			1455			1464			1473			1482	
ATC I	GGC G	CAT H	GAG E	CAG Q	GAG E	ACC T	ATG M	CTT L	GTG V	AAG K	ACA T	GCC A	AAA K	AAT N	GCT A	TAC Y	CAG Q
	1491			1500			1509			1518			1527			1536	
GTG V	GCA A	GGT G	GAG E	AGC S	CAA Q	GAG E	GAG E	GAG E	GGT G	CTA L	GAT D	GGA G	TAC Y	ACA T	GCT A	GTG V	GGA G
	1545			1554		;	1563			1572			1581			1590	
CCA P	CTT L	TTA L	GCT A	AGA R	AGG R	AAG K	Λ.	AGC S	ACC T	GGĀ G	GCC A	AGC S	AGC S	H	С	CAG Q	ĀĀG K
	1599			1608		:	1617	, 13,		1626			1635	( -	326) :	1644	
ACT T	TCT S	CTC L	AGG R	GTG V	AAC N	TTT F	GAG E	GAC D	ATC I	GGC G	TGG W	GAC D	AGC S	TGG W	ĀTC I	ĀTT I	GCA A
	1653		;	1662		;	L671		;	1680			1689			1698	
P P	AAG K	GAA E	TAT Y	GAC D	GCC A	TAT Y	GAG E	TGT C	ĀĀĀ K	GGG G	GGT G	TGC C	TTC F	TTC F	CCA P	TTG L	GCT A
	1707		;	1716		:	L725			1734			1743		1	L752	
GAT D	GAC D	GTG V	ACA T	CCC P	ACC T	ĀĀĀ K	CAT H	GCC A	ATC I	GTG V	CAG Q	ACC T	CTG L	GTG V	CAT H	CTC L	GAG E
	1761		;	2770		1	1779		:	1788			1797		3	1806	
TTC F	CCC P	ACA T	<del>AAG</del> K	GTG V	GGC G	AAA K	GCC A	TGC C	TGC C	GTT V	CCC P	ACC T	AAA K	CTG L	AGT S	CCC P	ATC I
;	1815		:	1824		1	833		:	1842		:	1851		1	.860	
TCC S	ATC I	CTC L	TAC Y	AAG K	GAT D	GAC D	ĀTG M	GGG G	GTG V	CCA P	ACC T	CTC L	AAG K	TAC Y	CAC H	TAT Y	GAG E
	1869			1878			.887					03		191	-		1923
GGG G	ATG M	AGI S	<u>GTG</u>	GCT A	GAG E	TGT C	GGG G	C .	AGG R 28)	TAGI	CCCT	GC 2	AGCCA	CCC	.G GG	TGGG	GATA

## Figure 1D

1933	1943	1953		1973	1983	1993
CAGGACATGG	AAGAGGTTCT	GGTACGGTCC	TGCATCCTCC	TGCGCATGGT	ATGCCTAAGT	TGATCAGAAA
2003	2013	2023	2033	2043	2053	2063
CCATCCTTGA	GAAGAAAAGG	AGTTAGTTGC	CCTTCTTGTG	TCTGGTGGGT	CCCTCTGCTG	AAGTGACAAT
2073	2083	2093	2103	2113	2123	2122
GACTGGGGTA	TGCGGGCCTG	TGGGCAGAGC	AGGAGACCCT	GGAAGGGTTA	GTGGGTAGAA	AGATGTCAAA
2143	2153	2163	2173	2183	2193	2203
AAGGAAGCTG	TGGGTAGATG	ACCTGCACTC	CAGTGATTAG	AAGTCCAGCC	TTACCTGTGA	GAGAGCTCCT
2213	2223	2233	2243	2253	2263	2773
GGCATCTAAG	AGAACTCTGC	TTCCTCATCA	TCCCCACCGA	CTTGTTCTTC	CTTGGGAGTG	TGTCCTCAGG
2283	2293	2303	2313	2323	2333	23/3
GAGAACAGCA	TTGCTGTTCC	TGTGCCTCAA	GCTCCCAGCT	GACTCTCCTG	TGGCTCATAG	GACTGAATGG
2353	2363	2373	2383	2393	2403	2413
GGTGAGGAAG	AGCCTGATGC	CCTCTGGCAA	TCAGAGCCCG	AAGGACTTCA	AAACATCTGG	ACAACTCTCA
242	3 243	3 244	13			3 - 2 - 2 - 2
TTGACTGAT	G CTCCAACAT	A ATTTTTAAA	A AGAG			

# Figure 2

10 20 30 40 50 60 70 CTCTAGAGGG CAGAGGAGGA GGGAGGGAGG GAAGGAGCG GGAGCCCGGC CCGGAAGCTA GGTGAGTGTG
80 90 100 110 120 130 140 GCATCCGAGC TGAGGGACGC GAGCCTGAGA CGCCGCTGCT GCTCCGGCTG AGTATCTAGC TTGTCTCCCC
150 160 170 180 190 200 210 GATGGGATTC CCGTCCAAGC TATCTCGAGC CTGCAGCGCC ACAGTCCCCG GCCCTCGCCC AGGTTCACTG
220 230 240 250 260 270 280 CAACCGTTCA GAGGTCCCCA GGAGCTGCTG CTGGCGAGCC CGCTACTGCA GGGACCTATG GAGCCATTCC
290 300 310 320 330 340 350 GTAGTGCCAT CCCGAGCAAC GCACTGCTGC AGCTTCCCTG AGCCTTTCCA GCAAGTTTGT TCAAGATTGG
360 370 380 390 400 (1) CTGTCAAGAA TCATGGACTG TTATTATATG CCTTGTTTTC TGTCAAGACA CC ATG ATT CCT MET Ile Pro
417 432 447 462 GGT AAC CGA ATG CTG ATG GTC GTT TTA TTA TGC CAA GTC CTG CTA GGA GGC GCG Gly Asn Arg MET Leu MET Val Val Leu Leu Cys Gln Val Leu Leu Gly Gly Ala
477 492 507 AGC CAT GCT AGT TTG ATA CCT GAG ACG GGG AAG AAA AAA GTC GCC GAG ATT CAG Ser His Ala Ser Leu Ile Pro Glu Thr Gly Lys Lys Val Ala Glu Ile Gln
522 537 552 567 GGC CAC GCG GGA GGA CGC CGC TCA GGG CAG AGC CAT GAG CTC CTG CGG GAC TTC Gly His Ala Gly Gly Arg Arg Ser Gly Glĥ Ser His Glu Leu Leu Arg Asp Phe
582 597 612 . 627 GAG GCG ACA CTT CTG CAG ATG TTT GGG CTG CGC CGC CCG CAG CCT AGC AAG Glu Ala Thr Leu Leu Gln MET Phe Gly Leu Arg Arg Pro Gln Pro Ser Lys
642 657 672 AGT GCC GTC ATT CCG GAC TAC ATG CGG GAT CTT TAC CGG CTT CAG TCT GGG GAG Ser Ala Val Ile Pro Asp Tyr MET Arg Asp Leu Tyr Arg Leu Gln Ser Gly Glu
687 702 717 732  GAG GAG GAA GAG CAG ATC CAC AGC ACT GGT CTT GAG TAT CCT GAG CGC CCG GCC Glu Glu Glu Glu Gln Ile His Ser Thr Gly Leu Glu Tyr Pro Glu Arg Pro Ala
747 762 777 AGC CGG GCC AAC ACC GTG AGG AGC TTC CAC CAC GAA GAA CAT CTG GAG AAC ATC Ser Arg Ala Asn Thr Val Arg Ser Phe His His Glu Glu His Leu Glu Asn Ile
792 807 822 837 CCA GGG ACC AGT GAA AAC TCT GCT TTT CGT TTC CTC TTT AAC CTC AGC AGC ATC Pro Gly Thr Ser Glu Asn Ser Ala Phe Arg Phe Leu Phe Asn Leu Ser Ser Ile

# Figure 2A

CCT Pro	GAG Glu	852 AAC Asn	GAG Glu	GTG Val	ATC Ile	TCC Ser	867 TCT Ser	GCA Ala	GAG Glu	CTT Leu	CGG Arg	882 CTC Leu	TTC Phe	CGG Arg	GAG Glu	CAG Gln	897 GTG Val
GAC Asp		GGC Gly	CCT Pro	912 GAT Asp	TGG Trp	GAA Glu	AGG Arg	GGC	927 TTC Phe	CAC His	CGT Arg	ATA Ile	AAC Asn	942 ATT Ile	TAT Tyr	GAG Glu	GTT Val
ATG MET	957 AAG Lys	CCC Pro	CCA Pro	GCA Ala	GAA Glu	972 GTG Val	GTG Val	CCT Pro	GGG Gly	CAC His	987 CTC Leu	ATC Ile	ACA Thr	CGA Arg		LOO2 CTG Leu	GAC Asp
	5	CTG	GTC Val	CAC His	CAC His	AAT Asn	GTG	1032 ACA Thr	CGG Arg	TGG Trp	GAA Glu	3.00	1047 TTT Phe	GAT Asp	GTG Val	AGC Ser	CCT Pro
106: GCG Ala	GTC Val		CGC Arg	TGG	1077 ACC Thr	CGG Arg	GAG Glu	AAG Lys	CNG	1092 CCA Pro	AAC Asn	TAT Tyr	GGG Gly	~~~	GCC Ala	ATT Ile	GAG Glu
GTG Val	ACT	CAC His	CTC Leu	CAT His	CAG Gln	ACT	CGG Arg	ACC Thr	CAC His	CAG Gln	GGC Gly	CAG Gln	CAT His	GTC Val	AGG Arg		AGC Ser
CGA Arg	TCG Ser	TTA Leu	CCT	CAA Gln	GGG Gly	AGT Ser	GGG Gly	A A T	197 TGG Trp	GCC Ala	CAG Gln	CTC Leu	000	CCC Pro	CTC Leu	CTG Leu	GTC Val
ACC	TTT Phe	GGC Gly	CAT His	GAT Asp	GGC	CGG Arg	GGC Gly	CAT His	GCC Ala	mmc	ACC Thr	CGA Arg	CGC Arg	CGG Arg		.272 GCC Ala	AAG Lys
CGT	AGC		.287 AAG	CAT	CAC	ጥሮኔ	CAG	L302	ccc	3.00	AAG		317				
Arg	Ser	Pro	Lys	His	His	Ser	Gln	Arg	Ala	Arg	Lys	Lys	AAT	AAG Lys	AAC	TGC Cys	Arg
CGC	(311 CAC His	TCG	CTC Leu	TAT	GTG Val	GAC Asp	TTC Phe	AGC Ser	CAT	.362 GTG Val	GGC Gly	TGG Trp	AAT Asn		.377 TGG Trp	ATT Ile	GTG Val
GCC Ala	CCA	392 CCA Pro	GGC Gly	TAC Tyr	CAG Gln	GCC	407 TTC Phe	TAC Tyr	TGC Cys	CAT His	GGG Gly	422 GAC Asp	TGC Cys	CCC Pro	TTT Phe		437 CTG Leu
GCT Ala	GAC Asp	CAC His	CTC	.452 AAC Asn	TCA Ser	ACC Thr	AAC Asn	CAT	.467 GCC Ala	ATT Ile	GTG Val	CAG Gln	100	.482 CTG Leu	GTC Val	AAT Asn	TCT Ser
GTC	497 AAT Asn	TCC Ser	AGT Ser	ATC Ile	CCC	512 AAA Lys	GCC Ala	TGT Cys	TGT Cys	GTG	.527 CCC Pro	ACT Thr	GAA Glu	CTG Leu	A CO	542 GCC Ala	ATC Ile

### Figure 2B

1557 1572 1587

TCC ATG CTG TAC CTG GAT GAG TAT GAT AAG GTG GTA CTG AAA AAT TAT CAG GAG
Ser MET Leu Tyr Leu Asp Glu Tyr Asp Lys Val Val Leu Lys Asn Tyr Gln Glu

1602 1617 (408) 1636 1646 1656 ATG GTA GAG GGA TGT GGG TGC CGC TGAGATCAGG CAGTCCTTGA GGATAGACAG MET Val Val Glu Gly Cys Gly Cys Arg

1736 1746 1756 1766 1776 1786 1796 ACAGACTGCT TCCTTATAGC TGGACTTTTA TTTAAAAAAA AAAAAAAAA AATGGAAAAA ATCCCTAAAC

1806 1816 1826 1836 1846 1856 1866 ATTCACCTTG ACCTTATTTA TGACTTTACG TGCAAATGTT TTGACCATAT TGATCATATA TTTTGACAAA

1876 1886 1896 1906 1916 1926 1936 ATATATTTAT AACTACGTAT TAAAAGAAAA AAATAAAATG AGTCATTATT TTAAAAAAAA AAAAAAAACT

1946 CTAGAGTCGA CGGAATTC



## Figure 3

_	ACA Thr -40	AGA Arg	GAG Glu	TGC Cys	TCA Ser	AGA Arg -35	AGC Ser	TGT Cys	CCA Pro	AGG Arg	ACG Thr -30	GCT Ala	CCA Pro	CAG Gln	AGG Arg	48
CAG Gln -25	GTG Val	AGA Arg	GCA Ala	GTC Val	ACG Thr -20	AGG Arg	AGG Arg	ACA Thr	CGG Arg	ATG Met -15	GCG Ala	CAC His	GTG Val	GCT Ala	GCG Ala -10	96
GGG Gly	TCG Ser	ACT Thr	TTA Leu	GCC Ala -5	AGG Arg	CGG Arg	AAA Lys	AGG Arg	AGC Ser 1	GCC Ala	GGG Gly	GCT Ala	GGC Gly 5	AGC Ser	CAC His	144
TGT Cys	CAA Gln	AAG Lys 10	ACC Thr	TCC Ser	CTG Leu	CGG Arg	GTA Val 15	AAC Asn	TTC Phe	GAG Glu	GAC Asp	ATC Ile 20	GGC Gly	TGG Trp	GAC Asp	192
AGC Ser	TGG Trp 25	ATC Ile	ATT Ile	GCA Ala	ccc Pro	AAG Lys 30	GAG Glu	TAT Tyr	GAA Glu	GCC Ala	TAC Tyr 35	GAG Glu	TGT Cys	AAG Lys	GGC Gly	240
GGC Gly 40	TGC Cys	TTC Phe	TTC Phe	ccc Pro	TTG Leu 45	GCT Ala	GAC Asp	GAT Asp	GTG Val	ACG Thr 50	CCG Pro	ACG Thr	AAA Lys	CAC His	GCT Ala 55	288
ATC Ile	GTG Val	CAG Gln	ACC Thr	CTG Leu 60	GTG Val	CAT His	CTC Leu	AAG Lys	TTC Phe 65	ĊCC Pro	ACA Thr	AAG Lys	GTG Val	GGC Gly 70	AAG Lys	336
GCC Ala	TGC Cys	TGT Cys	GTG Val 75	CCC Pro	ACC Thr	AAA Lys	CTG Leu	AGC Ser 80	CCC Pro	ATC Ile	TCC Ser	GTC Val	CTC Leu 85	TAC Tyr	AAG Lys	384
GAT Asp	GAC Asp	ATG Met 90	Gly	GTG Val	CCC Pro	ACC Thr	CTC Leu 95	AAG Lys	TAC Tyr	CAT His	TAC Tyr	GAG Glu 100	GGC Gly	ATG Met	AGC Ser	432
GTG Val	GCA Ala 105	GAG Glu	TGT Cys	GGG Gly	TGC Cys	AGG Arg 110	TAG	PATCI	rgc d	CTGC	GG					470

### INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 92/05374

I. CLASSIFI	CATION OF SUBJE	CT MATTER (If several classification :	symbols	apply, indicate all) <sup>6</sup>		
	5 C12N15/12	Classification (IPC) or to both National C 2; C12P21/02;	Classific	ation and IPC- A61K37/02		
II. FIELDS	SEARCHED					
		Minimum Docum	nentatio	n Searched <sup>7</sup>		
Classification	on System		Classi	fication Symbols		
Int.Cl.	5	CO7K ; C12N ;		A61K		
		Documentation Searched other to the Extent that such Documents	er than l	Minimum Documentation cluded in the Fields Searched <sup>8</sup>		
III. DOCU	MENTS CONSIDER	ED TO BE RELEVANT <sup>9</sup>		12	Relevant to Claim No.13	
Category "	Citation of D	ocument, 11 with indication, where approp	priate, o	f the relevant passages w	RECYALL TO CLAIM 1407	
A	4 Octob	011 366 (GENETICS INST per 1990 n the application	TITUT	TE, INC.)	1-18	
A	see the	1-18				
	SCIENCE vol. 87 US pages 9 CELESTE transfo member: purific see the					
				-/		
° Spec	ial categories of cited	documents: 10		later document published after the or priority date and not in conflict	ALLY IDS TRUITSTING ON!	
TE" e	"A" document defining the general state of the art which is not considered to be of particular relevance "E" earlier document but published on or after the international filing date "I." document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified) "O" document referring to an oral disclosure, use, exhibition or other means "P" document published prior to the international filing date but later than the priority date claimed  "A" document of particular relevance; the cannot be considered novel or cannot be cannot be considered novel or cannot be cannot be considered to involve an invo					
	TIFICATION		<del></del> -	Date of Mailing of this Internation	al Search Report	
Date of the		of the International Search TOBER 1992		19. 10. 9°		
Internatio	onal Searching Author	ITY		Signature of Authorized Officer ANDRES S.M.		

III. DOCUMENTS CONSIDERED TO BE RELEVANT (CONTINUED FROM THE SECOND SHEET)								
Category °	Citation of Document, with indication, where appropriate, of the relevant passages	Relevant to Claim No.						
P,A	WO,A,9 118 098 (GENETICS INSTITUTE, INC.) 28 November 1991 cited in the application see the whole document	1-18						

#### INTERNATIONAL SEARCH REPORT

Leternational application No.

PCT/US 92/05374

Rox	1 Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)	
This	international search report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:	
1. [	Claims Nos.: because they relate to subject matter not required to be searched by this Authority, namely:  Remark: Although claims 16, 18 are directed to a method of treatment of the human/animal body the search has been carried out and based on the alleged affects of the compound/composition.	
2. [	Claims Nos.: because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:	
3. [	Claims Nos.: because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).	
Box	x II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)	7
Thi	s International Searching Authority found multiple inventions in this international application, as follows:	
1.	As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.	
2.	As all searchable claims could be searches without effort justifying an additional fee, this Authority did not invite payment of any additional fee.	
3.	As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:	
4.	No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:	
R	emark on Protest  The additional search fees were accompanied by the applicant's protest.  No protest accompanied the payment of additional search fees.	

#### ANNEX TO THE INTERNATIONAL SEARCH REPORT ON INTERNATIONAL PATENT APPLICATION NO. US 9205374 61850

This armer lists the patent family members relating to the patent documents cited in the above-mentioned international search report. The members are as contained in the European Patent Office EDP file on The European Patent Office is in no way liable for these particulars which are merely given for the purpose of information. 05/10/92

Patent document cited in search report	Publication date	F	Patent family member(s)	Publication date	
WO-A-9011366	04-10-90	US-A- AU-A- CA-A- EP-A- JP-T-	5106748 5357790 2030518 0429570 3505098	21-04-92 22-10-90 29-09-90 05-06-91 07-11-91	
√0-A-9118098	28-11-91	None			
				•	

FORM POOP

ੇ For more details about this annex : see Official Journal of the European Patent Office, No. 12/82